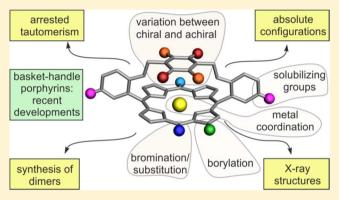
Monomeric Chiral and Achiral Basket-Handle Porphyrins: Synthesis, Structural Features, and Arrested Tautomerism

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Supporting Information

ABSTRACT: Chiral and achiral basket-handle porphyrins (BHPs) with different *p*-xylene straps and peripheral solubilizing groups were synthesized using a previously established synthetic approach. Subsequent modification, functionalization, and metalation of the tetrapyrrolic macrocycle yielded more than 80 BHPs. The chiral representatives were resolved into their enantiomers, whose absolute configurations were determined by comparison of their ECD spectra with other experimental or quantum chemically calculated spectra. NMR studies and coupled-cluster calculations proved that the free base BHPs, although highly symmetric, exhibited the phenomenon of "arrested tautomerism". Comparison of the solid-state structures of three metalated BHPs offered detailed insight into their three-

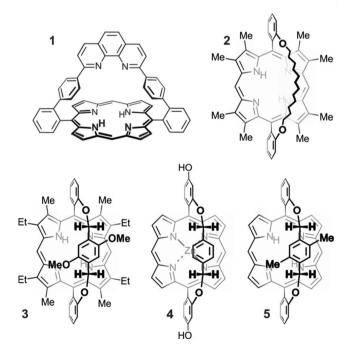


dimensional shape. Finally, directly linked dimeric porphyrins with a BHP subunit were synthesized from functionalized BHPs to prove their value as synthetic building blocks.

INTRODUCTION

Porphyrins are one of the most versatile chemical structures and therefore have been widely investigated since they not only combine interesting optical, photophysical, and catalytic properties but also allow these properties to be tailored to particular needs.¹ This facilitates their use in almost all fields of natural sciences, such as medicine,² biology,³ physics,⁴ and particularly in chemistry.¹

Within the field of chemistry, the construction of defined three-dimensional geometries of either monomeric tetrapyrroles⁵ or arrays of multiple systems^{6,7} has been a central topic, especially during the past few decades. This required suitable strategies for the construction, modification, and combination of the building blocks. Porphyrins with steric shielding on one or both sides of the macrocyclic plane, known as capped or basket-handle porphyrins⁸⁻¹² (BHPs, examples 1–5), although known for a long time, have however not attained a pronounced impact.^{13,14} This was mainly due to the difficult syntheses of such systems, which often involved multiple steps⁸ and/or low yields^{15–19} and their unfavorable properties such as chemical instability, poor solubility,¹⁶ or lack of possibilities for subsequent modification. Just recently we have described a synthetic approach that overcomes these difficulties and gives access to multigram quantities of BHPs. Both chiral (5) and achiral forms have thus become available, and these can also be further modified.¹²



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There is one example of a related system in the literature, compound 3, for which we expect the same chirality as in our systems, but this was not discussed at the time.¹⁰

This work gives extensive insight into the versatility of the synthesis of the BHPs, their further modification, and their use as synthetic building blocks. Furthermore, we will present detailed structural investigations, especially regarding their planar chirality and arrested tautomerism.

RESULTS AND DISCUSSION

In our previous report on 5,15-bridged BHPs¹² we introduced a short and efficient synthesis of the BHPs and gave the first examples of their modification. The various sites available for modification are summarized in a structural representation (Figure 1). The following sections will provide detailed insight regarding each of the modification sites.

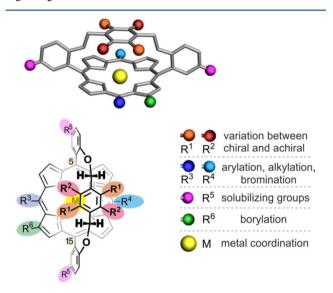


Figure 1. Possible sites for the modification of the 5,15-bridged BHPs (adapted from ref 12, copyright 2015, American Chemical Society).

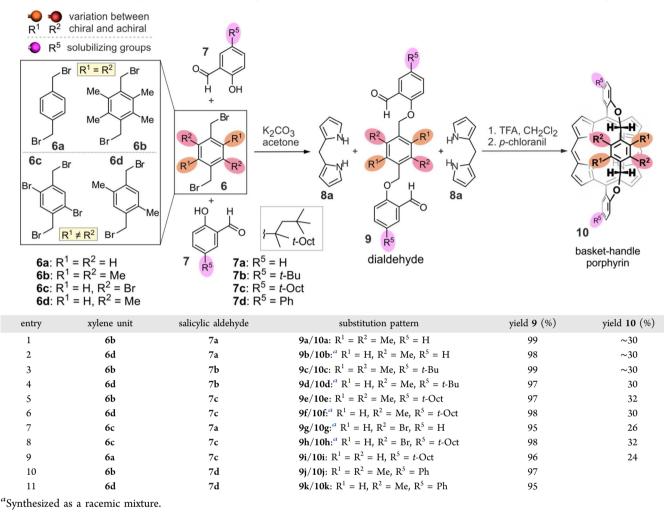
Variation between Chiral and Achiral Forms and Introduction of Solubilizing Groups. The synthetic approach to the BHPs starts with the construction of a suitable *p*-xylene-linked²⁰ dialdehyde 9, which reacts with dipyrrylmethane 8 to yield the BHPs 10 after a two-step, one-pot reaction consisting of a cyclocondensation and a subsequent oxidation of the initially formed porphyrinogen (Table 1). This strategy requires the careful selection of the appropriate dialdehyde 9, since the decision between a later planar-chiral, C_2 -symmetric BHP or an achiral, $C_{2\nu}$ -symmetric BHP has to be made at the stage of the dialdehyde synthesis. By a choice of handle derivatives $6a_{,b}$ with four identical substituents (R^1 = R^2), the resulting BHPs have $C_{2\nu}$ symmetry and are therefore achiral. If the substituents along the diagonal of the xylene unit are not identical $(R^1 \neq R^2)$, as in **6c,d**, the porphyrin formation yields a racemic mixture of two planar-chiral enantiomers. A total of four achiral BHPs (10a,c,e,i) with either four methyl groups or four hydrogens within the xylene ring were synthesized by this strategy. Likewise, five planar-chiral BHPs (10b,d,f-h) were prepared with either two bromine or two methyl substituents along the diagonal of the xylene ring and two hydrogens at the remaining positions.

The second site open to modification prior to the porphyrin formation is the position para to the hydroxy function in the salicylic aldehydes 7, used for the preparation of the *p*-xylenelinked dialdehyde 9. This position is the ideal site for solubilizing groups for two reasons. First, the position para to the hydroxy function is the most remote with regard to the porphyrin core and the basket handle in the final BHP and therefore has the least influence on the BHP properties. Second, the required substituted salicylic aldehydes 7 are easily prepared by literature procedures.^{21,22} The parent compound salicylic aldehyde 7a eventually gives a BHP with no solubilizing substituent yet is still sufficiently soluble for most applications. Introduction of a tert-butyl group slightly increases the solubility, while a tert-octyl substituent improves it significantly. These findings were expected and were in agreement with factors usually mentioned when discussing the solvation of porphyrins.²³ As will be shown later, in sterically controlled Ir-catalyzed C-H activation reactions,² these alkyl substituents are sterically necessary to prohibit C-H activation at the remaining phenyl positions. Interestingly, the phenyl-substituted dialdehydes 9j,k did not yield any porphyrin when subjected to the otherwise successful condensation conditions. We currently have no hints as to whether this is due to solubility problems at the porphyrinogen or porphyrin stage or whether it is a problem caused by a different reactivities of the phenyl-substituted dialdehyde. However, in summary, the systematic choice of suitable salicylic aldehydes and pxylene units permits the directed synthesis of BHPs.

Synthesis of *meso*-Substituted BHPs by Mixed Condensation or by "Senge Reaction" with Lithium Organyls. Although 5,15-disubstituted porphyrins with two remaining free *meso*-positions can easily be prepared²⁵ and have frequently been investigated, their closely related 5,10,15-triand 5,10,15,20-tetrasubstituted analogues might be of even greater interest.^{5,26} Therefore, we adopted two of our protocols to the construction of such systems to our BHPs. First, altering the parameters at the porphyrin-forming step, we exchanged half of the amount of the unsubstituted dipyrrylmethane **8a** with substituted species (**8b,c**), which resulted in a setup usually called "mixed condensation" (Table 2).²⁶ As expected, we obtained a statistical mixture of the respective di-, tri-, and tetrasubstituted BHPs.

If only the trisubstituted BHP **11** was the desired product and there was no use for the di- and tetrasubstituted side products **10** and **12**, a more direct approach was favorable. As we previously reported, the method developed by Senge,²⁷ which uses lithium organyls for the selective *meso*-substitution of initially *meso*-unsubstituted porphyrins, can also be applied to our BHPs. So far only achiral phenyl- and *n*-butyl-substituted BHPs (entries 8 and 9, BHPs **11e**,**f**) without solubilizing groups have been synthesized by this strategy.¹² We have expanded this protocol to further alkyl and aryl substituents and applied it to several BHPs with solubilizing groups, but also to a chiral species (Table 2).

meso-Bromination and Debromination of the BHPs. Our previous work on the bromination of BHPs only included the full *meso*-bromination of the simplest chiral and achiral BHPs **10a**,**b**.¹² Herein we report the full scope of this reaction. Regardless of the substitution pattern of the handle or the presence of solubilizing groups, all BHPs with two free *meso*positions **10a**–**h** were fully *meso*-brominated using 2.1 equiv of NBS in chloroform. Furthermore, aryl- and alkyl-substituted BHPs **11b**,**e**,**h** with only one free *meso*-position were brominated with 1.05 equiv of NBS in almost quantitative yields. Finally, we synthesized the monobrominated BHPs Table 1. Variation of the p-Xylene Unit Leading to Chiral or Achiral BHPs (10) and Introduction of Solubilizing Groups

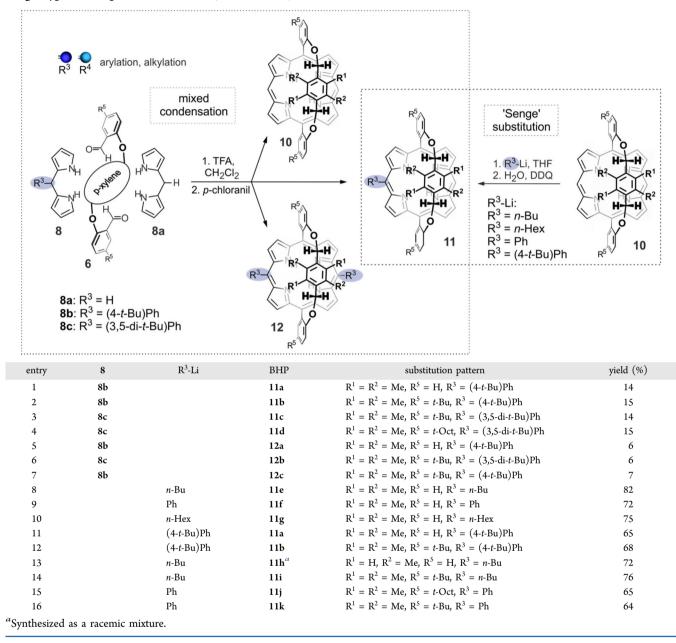


14d,e with one remaining free *meso*-position (Table 3). Since bromination of porphyrins with more than one free *meso*position is not selective,²⁸ the bromination of 10a,c yielded a mixture of mono- and dibrominated BHPs along with the unreacted BHP. The highest yields of monobrominated BHPs 14d,e were obtained with 1.2 equiv of NBS, and the reaction mixture was easily resolved by column chromatography. As previously reported, it was essential to carry out all bromination reactions without pyridine, as this led to decomposition.¹² The reverse reaction, the hydrodebromination of brominated porphyrins via a method described by Osuka et al. using Pd(PPh₃)₄ as a catalyst and formic acid as a hydrogen source,²⁹ was successfully applied to the brominated BHP 13c.

Highly Regioselective Ir-Catalyzed β -Borylation of Porphyrins by C–H Bond Activation. The previous report on the direct borylation of BHPs via Ir-catalyzed C–H activation was limited to the 5,15-bridged BHP 10c, possessing two remaining free *meso*-positions.¹² However, the protocol was found not to be suitable for the β -borylation of *meso*-phenylsubstituted porphyrins, since C–H activation at the phenyl ring occurred as an undesired side reaction.³⁰ Instead, the direct borylation of BHP 11b with 3,5-di-*tert*-butylphenyl substitution in one *meso*-position and *tert*-butyl groups at the basket-handle aryls seemed more promising (Table 4). The *tert*-butyl groups at the *meso*-phenyl substituent and the *meso*-bridge phenyls were essential for the regioselectivity of the reaction, since without them borylation of any sterically unhindered phenyl group was possible. Therefore, the *tert*-butyl groups at the *meso*bridge phenyls not only increased solubility but also acted as directing groups in the sterically controlled C–H activation reactions. Depending on the amount of the pinacolborane source—either HBpin or $(BPin)_2$ —the yield was optimized to give favorably diborylated BHP **15g** in 77% yield with an excess of the borylation reagent or monoborylated BHP **15f** in 55% yield using 2 equiv of the borylating agent. Unreacted starting material and the two products were easily resolved by recycling gel-permeation chromatography.

Suzuki Coupling with BHP Building Blocks. To demonstrate the applicability of functionalized BHPs as synthetic building blocks, we synthesized two directly linked dimeric porphyrins, each containing a BHP unit (Scheme 1). Both dimers were synthesized by Suzuki coupling under conditions reported previously.³¹ Dimer 18 was formed from the β -brominated tetraarylporphyrin 16a and the β -borylated BHP 15a in a yield of 83%. Miyaura borylation³² of 16b yielded the β -borylated tetraarylporphyrin 17, which was coupled with *meso*-brominated BHP 14e to give the dimer 19 (78% yield). Both dimers were found to adopt a conformation with the *meso*-aryl substituent of the tetraarylporphyrin unit located on the sterically unhindered side of the BHP. Investigations regarding the stereostructures and chirality of these dimers are in progress and will be reported later.

Table 2. *meso*-Substituted BHPs by Mixed Condensation (Entries 1–7) with 5-Substituted Dipyrrylmethanes (8b,c) or by Senge-Type Nucleophilic Substitution (Entries 8–16)

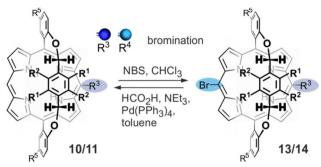


Metalation of BHPs. Previously we described a series of achiral BHPs which contained different metals in the oxidation state +2, namely Pd, Ni, Zn, Cu, and Mg.¹² We were able to expand the scope of metallo-BHPs to oxidation states higher than +2 by synthesizing BHPs with In(III) and V(IV) as complexed metals in quantitative yield (Table 5). The remaining free axial coordination site is occupied by chloride for all reported In(III) BHPs and by an oxo ion for the V(IV) BHP. As already described earlier for most of the M^{II}-BHPs, demetalation by acidic, basic, or reductive conditions was not possible for the ions in higher oxidation states, which is due to the steric shielding of the handle that prevents the metal ion from being expelled from the porphyrin.¹²

Solid-State Structures and Conformations in Solution. Previous results of NMR investigations and computational structure optimizations gave first insight into the stereostructures of BHPs. Additional information about the threedimensional shape of a metalated *p*-xylene-strapped BHP was obtained by the crystal structure of Ni-10a.¹² By slow diffusion of methanol (containing 5% water) into a dilute solution of the respective BHP in chloroform, we succeeded in growing crystals of V-10d and In-10d suitable for X-ray diffraction investigations. These results (Figure 2) in combination with DFT-optimized structures were used to get a more detailed insight into the conformational differences of the BHPs.

The solid-state structures of Ni-10a, In-10d, and V-10d showed most of the possible conformations that the BHPs can adopt. Ni-10a and V-10d possessed a large gap between the porphyrin macrocycle (defined by the four nitrogen atoms) and the bridge (using a centroid of the carbons of the xylene ring) with a distance above 4.70 Å, while this gap was significantly smaller in In-10d (3.50 Å). Another difference was found when comparing the angles between the planes of the xylene unit and tetrapyrrole. While these planes were nearly coplanar for In-

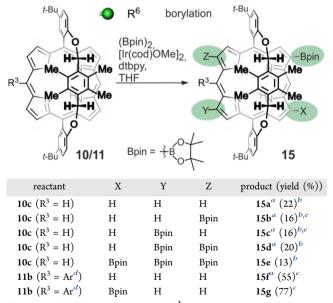
Table 3. Bromination (Entries 1-14) and Debromination (Entry 15) of BHPs



entry	reactant	reagent	product	yield (%)
1	10a: $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = H$	NBS (2.1 equiv)	13a: $R^1 = Me$, $R^2 = Me$, $R^3 = Br$, $R^5 = H$	98
2	10b : $R^1 = H$, $R^2 = Me$, $R^3 = H$, $R^5 = H$	NBS (2.1 equiv)	13b : ^{<i>a</i>} $R^1 = H$, $R^2 = Me$, $R^3 = Br$, $R^5 = H$	98
3	10c: $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = t$ -Bu	NBS (2.1 equiv)	13c: $R^1 = Me$, $R^2 = Me$, $R^3 = Br$, $R^5 = t$ -Bu	97
4	10d: $R^1 = H$, $R^2 = Me$, $R^3 = H$, $R^5 = t$ -Bu	NBS (2.1 equiv)	13d : ^{<i>a</i>} $R^1 = H$, $R^2 = Me$, $R^3 = Br$, $R^5 = t$ -Bu	99
5	10e : $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = t$ -Oct	NBS (2.1 equiv)	13e : $R^1 = Me$, $R^2 = Me$, $R^3 = Br$, $R^5 = t$ -Oct	97
6	10f: $R^1 = H$, $R^2 = Me$, $R^3 = H$, $R^5 = t$ -Oct	NBS (2.1 equiv)	13f : ^{<i>a</i>} $R^1 = H$, $R^2 = Me$, $R^3 = Br$, $R^5 = t$ -Oct	96
7	10g: $R^1 = H$, $R^2 = Br$, $R^3 = H$, $R^5 = H$	NBS (2.1 equiv)	13g: ^{<i>a</i>} $R^1 = H$, $R^2 = Br$, $R^3 = Br$, $R^5 = H$	97
8	10h : $R^1 = H$, $R^2 = Br$, $R^3 = H$, $R^5 = t$ -Oct	NBS (2.1 equiv)	13h : ^{<i>a</i>} $R^1 = H$, $R^2 = Br$, $R^3 = Br$, $R^5 = t$ -Oct	99
9	10i: $R^1 = H$, $R^2 = H$, $R^3 = H$, $R^5 = t$ -Oct	NBS (2.1 equiv)	13h : $R^1 = H$, $R^2 = H$, $R^3 = Br$, $R^5 = t$ -Oct	99
10	11h: $R^1 = H$, $R^2 = Me$, $R^3 = n$ -Bu, $R^5 = H$	NBS (1.05 equiv)	14a: ^{<i>a</i>} $R^1 = H$, $R^2 = Me$, $R^3 = n$ -Bu, $R^5 = H$	96
11	11e: $R^1 = Me$, $R^2 = Me$, $R^3 = n$ -Bu, $R^5 = H$	NBS (1.05 equiv)	14b : $R^1 = Me$, $R^2 = Me$, $R^3 = n$ -Bu, $R^5 = H$	94
12	11b : $R^1 = Me$, $R^2 = Me$, $R^3 = (4-t-Bu)Ph$, $R^5 = t-Bu$	NBS (1.05 equiv)	14c : $R^1 = Me$, $R^2 = Me$, $R^3 = (4-t-Bu)Ph$, $R^5 = t-Bu$	99
13	10c : $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = t$ -Bu	NBS (1.2 equiv)	14d : $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = t$ -Bu	62
14	10a : $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = H$	NBS (1.2 equiv)	14e : $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = H$	59
15	13c : $R^1 = Me$, $R^2 = Me$, $R^3 = Br$, $R^5 = t$ -Bu	$HCO_{2}H$, NEt ₃ , Pd(PPh ₃) ₄	10c : $R^1 = Me$, $R^2 = Me$, $R^3 = H$, $R^5 = t$ -Bu	91

^{*a*}Synthesized as a racemic mixture.

Table 4. Direct Borylation by Ir-Catalyzed C-H Activation



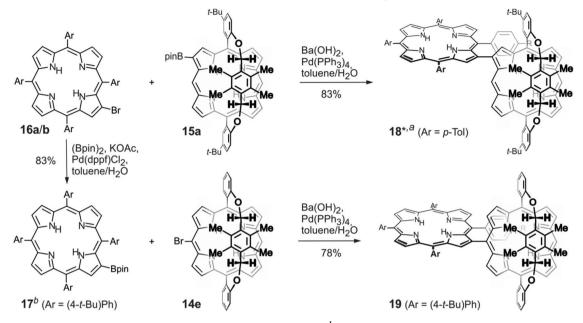
^{*a*}Synthesized as a racemic mixture. ^{*b*}Statistical product distribution, not optimized for a specific product. ^{*c*}Optimized yield. ^{*d*}Ar = 3,5-*t*-Bu₂Ph. ^{*e*}Inseparable mixture of the two regioisomers.

10d, they were strongly tilted for the metalated BHPs with large gaps $(44^{\circ} \text{ for Ni-10a and } 33^{\circ} \text{ for V-10d})$.

For a more systematic investigation of the conformational behavior of the BHPs, we performed conformational analyses of Ni-10, Zn-10, and 2H-10, with either two or four methyls in the strap, using B97-D3/def2-TZVP. Surprisingly, these analyses all yielded only one possible conformation according to a Boltzmann statistical weighting. All achiral BHPs with a four-methyl strap (M-10a and M-10c) showed a large gap between the bridge and the porphyrin, while the chiral BHPs with only two methyls (M-10d) in the strap had a short distance. This was easily observed by comparing the dihedral angles C-1'-C-2'-O-3'-C-4', which were nearly 0° in cases containing the tetramethylated bridge and nearly 90° for the dimethylated species. In addition, the methylene protons at the 4'-position of M-10d were always oriented toward the methyl group of the xylene unit, while the conformation with the methylene protons directed toward the hydrogen of the xylene unit was in all cases significantly higher in energy and should not be observable in solution. The reason for the different gaps is the interplay of steric hindrance (especially of the methyl groups) with the stabilizing dispersion effects (e.g., $\pi - \pi$ stacking) between the bridge and the porphyrin plane. The steric hindrance with four methyl groups was obviously higher than the stabilizing effect of the $\pi - \pi$ stacking in 2H-10c, Zn-10a, and Ni-10a, and thus a maximum distance between the bridge and the porphyrin was reached. For the chiral BHPs with only two methyls the stabilization of the molecule by the π - π stacking became stronger than the steric hindrance and the distance between strap and porphyrin plane became smaller.

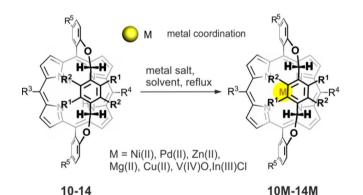
NMR investigations clearly confirmed the findings discussed above for the solid-state structures and the calculated conformations. The chiral BHPs (with two methyl or two bromine substituents in the xylene unit) show distinct differences in their chemical shifts and symmetry in comparison

Scheme 1. Synthesis of Directly Linked Dimeric Porphyrins from BHP Building Blocks



^aSynthesized as a racemic mixture; 18 was synthesized from 16a (Ar = p-Tol). ^b17 was prepared from 16b.

Table 5. Metalation of BHPs



			10-14		10141-14141		
reactant	\mathbb{R}^1	\mathbb{R}^2	R ³	\mathbb{R}^4	R ⁵	product	М
10a	Me	Me	Н	Н	Н	M-10a	Ni, ^b Pd, ^c Zn, ^d Mg, ^e Cu ^f
10b	Me	Н	Н	Н	Н	M-10b ^a	Ni, Zn, In ^g
10c	Me	Me	Н	Н	<i>t</i> -Bu	M-10c	Pd, In, Zn, Ni
10d	Me	Н	Н	Н	<i>t</i> -Bu	M-10d ^a	Ni, Pd, Zn, Mg, Cu, In, V^h
10e	Me	Me	Н	Н	<i>t</i> -Oct	M-10e	Zn
10f	Me	Н	Н	Н	<i>t</i> -Oct	M-10f ^a	In
10h	Br	Н	Н	Н	<i>t</i> -Oct	M-10h ^a	In
10i	Н	Н	Н	Н	<i>t</i> -Oct	M-10i	Ni
11a	Me	Me	Н	(4-t-Bu)Ph	Н	M-11a	Zn, Cu, Ni
11b	Me	Me	Н	(4-t-Bu)Ph	<i>t</i> -Bu	M-11b	Pd, Ni, Cu, Zn
11e	Me	Me	Н	<i>n</i> -Bu	Н	M-11e	Zn, Ni
11i	Me	Me	Н	<i>n</i> -Bu	<i>t</i> -Bu	M-11i	Zn
11h	Me	Н	Н	<i>n</i> -Bu	Н	M-11h ^a	Zn
11g	Me	Me	Н	<i>n</i> -Hex	Н	M-11g	Zn, Ni
12a	Me	Me	(4- <i>t</i> -Bu)Ph	(4-t-Bu)Ph	Н	M-12a	Zn, Ni
13h	Br	Н	Br	Br	<i>t</i> -Oct	M-13h ^a	In
13f	Me	Н	Br	Br	<i>t</i> -Oct	M-13f ^a	In
13c	Me	Me	Br	Br	<i>t</i> -Bu	M-13c	Ni
14d	Me	Me	Br	Н	<i>t</i> -Bu	M-14d	Ni

^{*a*}Synthesized as a racemic mixture. ^{*b*}Ni(acac)₂, (CH₂Cl)₂, ΔT . ^{*c*}Pd(OAc)₂, (CH₂Cl)₂, ΔT . ^{*d*}Zn(OAc)₂, CHCl₃, ΔT . ^{*e*}MgBr₂·OEt₂, NEt₃, CHCl₃, ΔT . ^{*f*}Cu(OAc)₂, CHCl₃, ΔT . ^{*f*}MgBr₂·OEt₂, NEt₃, CHCl₃, ΔT .

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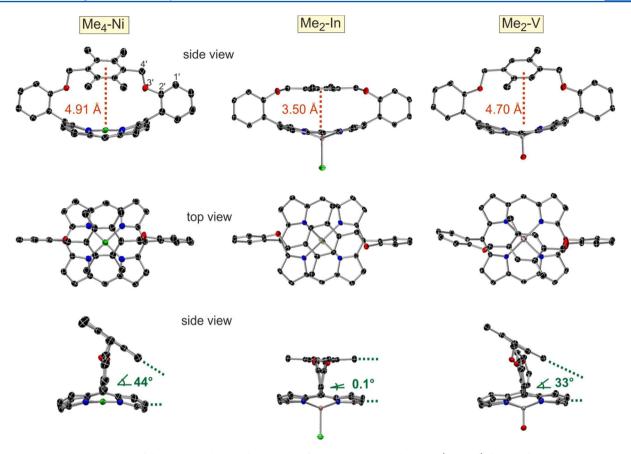


Figure 2. ORTEP representations of the X-ray single crystal structures of Ni-10a, In-10d, and V-10d: (top, red) distance between the centroid of the pyrrole nitrogens and the centroid of the aromatic xylene carbons; (bottom, green) angle between the xylene plane and the tetrapyrrolic plane. All disorder, hydrogens bonded to carbon, and peripheral *tert*-butyl groups removed for clarity.

to the achiral BHPs (with four methyl substituents in the xylene unit). The two protons of the methylene unit as well as the methyl groups and the hydrogens of the xylene ring represent ideal diagnostic groups. For the achiral BHPs the two methylene protons appeared as a sharp singlet, thus being magnetically identical, and showed no interaction with the pyrrolic protons in ¹H NOE experiments. This proved a $C_{2\nu}$ symmetric conformation in solution on the NMR time scale. The sharp shape of the signal of the methylene protons (as well as for all protons within the strap) indicated a rigid conformation, as changes in the location of the strap unit relative to the porphyrin macrocycle would induce peak broadening due to an altering position of the protons within the porphyrin ring current. In contrast, each of the two methylene protons of the chiral BHPs appeared as a sharp doublet with largely different chemical shifts (Figure 3), proving a C_2 -symmetric conformation in solution. Both protons showed a strong interaction with the methyl group of the strap, while only one of these protons showed a ¹H NOE interaction with only one pyrrole unit (see the Supporting Information for NOE spectra), and none of the methylene protons interacted with a proton of the xylene ring. This proved that in solution the solely present conformation has the methylene protons directed at the methyl group, which confirmed the assumptions from the calculations. Furthermore, we compared the ¹H chemical shifts of the methylene protons and the methyl groups between various BHPs (Figure 3) which differ only in the number of methyl groups in the strap and the coordinated metal ion. The results showed that the chemical shift of these

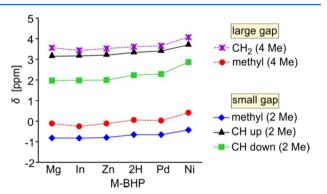


Figure 3. 1 H NMR shifts of the benzylic xylene protons and the xylene methyl groups of chiral (M-10d) and achiral (M-10c) BHPs.

groups varied largely between the chiral and the achiral BHPs and this difference correlated with the distance between the xylene unit and the porphyrin and was caused by the ring current of the porphyrin, which is inversely proportional to the distance to the porphyrin. These results not only substantiated the discussed differences between the chiral and achiral BHPs but also proved that the conformations among only chiral or only achiral porphyrins are very similar.

Due to the open-shell nature of the coordinated V(IV) ion, it was not possible to acquire NMR data for **V-10d** that would provide information about its conformation in solution. To find the reason **V-10d** showed significant deviations in the crystal structure in comparison to the other BHPs with dimethylated bridges, we performed a conformational analysis of **V-10d** using

B97D3/def2-TZVP. In contrast with the results described above, where solid-state structures and conformations derived from the calculations and from NMR investigations were in accordance, the solid-state structure and the computed minmum of V-10d differed clearly, as the DFT analysis predicted the same conformation for the vanadium BHP as for all other chiral BHPs: i.e., with a small gap between the strap and the porphyrin. To exclude that this is an artifact from the method used, single-point energy calculations were done using B3LYP-D3, PW6B95-D3, and SCS-CC2 (all with the def2-TZVP basis set), and all methods confirmed the previous results (the large-gap structure is about 11 kcal/mol higher in energy). Another hint at the conformation in solution was obtained by ECD investigations. The calculated ECD spectra of small-gap V-10d (see the Supporting Information) showed a much better fit with the experimental curves than the conformation with the large gap. Thus, most probably, the deviations in the X-ray structure are due to only packing effects.

Assignment of the Absolute Configuration of Metalated BHPs. We previously reported the absolute stereostructure of Ni-10b, which we obtained by comparing online ECD spectra with calculated spectra.¹² Measurements were done after resolution of the enantiomers by HPLC on a chiral phase. Using the same approach, we succeeded in resolving the enantiomers of all metallo-BHPs M-10d on a Chiralpak IA column using isocratic solvent mixtures (for chromatographic details, see the Supporting Information) and measured the online ECD spectra of the enantiomers. The faster-eluting enantiomers always displayed a negative couplet within the Soret band region while, vice versa, the slower-eluting enantiomers showed a positive, mirror-image couplet (Figure 4). All ECD spectra were very similar, regardless of the central metal, even for open-shell ions such as Cu(II) and V(IV). As

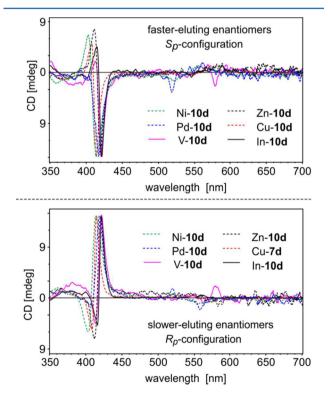


Figure 4. Experimental ECD spectra of (top) S_p -M-10d and (bottom) R_p -M-10d: online measurements, scaled to the most intense signal.

previous investigations of the absolute configurations of porphyrins with different kinds of chirality (stereocenters,³³ helical chirality,^{34,35} axial chirality^{36,37}) had shown that the metal centers do not influence the ECD spectra significantly (e.g., sign inversion had never been observed), the absolute configurations of the metalated BHPs were determined by simple comparison of the experimental ECD spectra. Thus, as all faster-eluting enantiomers show a negative ECD signal at ~420 nm, they do have the S_p configuration, while all slower-eluting species are R_p -configured.

To verify the previous determination of the absolute configurations of the enantiomers of Ni-10b,¹² we compared the results of TD B3LYP, TD BHLYP, and TD CAM-B3LYP calculations of Ni-10b and Zn-10b with those of SCS-CC2. Using a UV shift of 38 nm, the SCS-CC2-predicted ECD spectra for Zn-10b showed a nice fit with the experimental results and confirmed the previously published results for Ni-10b.¹² CAM-B3LYP and BHLYP gave nearly identical results (except for different UV shifts), and thus BHLYP could be neglected in the following. Both functionals, B3LYP and CAM-B3LYP, permitted a correct determination of the absolute configuration (see Figure S1 in the Supporting Information) of the investigated BHPs with Δ_{ESI} values³⁸ above 85% for CAM-B3LYP (0.08 eV, 48 nm shift) and above 70% for B3LYP (0.08 eV, 30 nm shift). However, the CAM-B3LYP ECD curves showed a much better fit with the experimental spectra and the SCS-CC2 results than the B3LYP curves. The reasons for this were several wrongly predicted CT states (ghost states³⁹) in the B3LYP spectra, which falsified the results. All of these states had an electron-hole distance D^{40} above 2.7 Å and an overlap S^{41} of less than 10%, clearly confirming that B3LYP cannot correctly handle these excitations. The SCS-CC2-predicted ECD curves (shift of 58 nm) of Ni-10b fit very well to the measured spectra, and again CAM-B3LYP and BHLYP showed comparable spectra while the match of the B3LYP results was significantly worse. In the B3LYP calculations a high number of excitations from the d_{z^2} to the $d_{x^2-y^2}$ orbital of the nickel atom occurred. These were only partially observed in the SCS-CC2 computations. In addition, several ghost states below the Soret band region showed wrong ECD signs in comparison to the results of the experiment or the other calculations (selected electron hole distances D and overlaps S for the TDDFT results can be found in Table S3 in the Supporting Information). In general, TDB3LYP was found to seriously suffer from ghost states when the excited states of BHPs were calculated and thus CAMB3LYP or BHLYP should be preferred for these investigations.

Assignment of the Absolute Configuration and Investigation of the Tautomerism of the Chiral Free-Base BHPs. The enantiomers of the chiral free-base BHPs were—like those of the metaled chiral BHPs—resolvable by HPLC on a chiral phase. However, to our surprise the online CD measurements gave ECD spectra that were not comparable with those of the metalated BHPs. Initially we attributed this effect to be related to the NH tautomerism, a characteristic of porphyrins, which has been known for over half a century and has been studied extensively.^{42–44} Our chiral free base BHPs, which are C_2 symmetric, were synthesized as racemic mixtures, with each enantiomer potentially existing as a mixture of two tautomers. These tautomers are, at the same time, diastereomers (Figure 5).

We expected a strong influence of the tautomerism on the ECD spectra. Indeed, a comparison of the calculated curves of

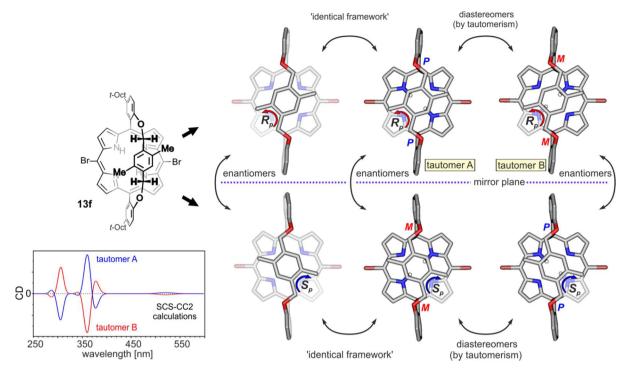


Figure 5. Tautomerism in chiral BHPs. tert-Octyl substituents and selected protons are omitted for clarity.

the tautomers showed that they are nearly mirror images (Figure 5). Thus, the Boltzmann statistical weighting of the single curves had to be as precise as possible to get reliable ECD spectra, and several methods were evaluated to obtain the relative energies of the tautomers of 13f. The initially used optimization method B97D3/def2-TZVP preferred the tautomer with the hydrogens orthogonal to the bridge methyl groups (tautomer A, Figure 5) only by 0.88 kcal/mol (ΔE). In the following, tautomer A is always the reference compound, so that positive relative energies refer to a preference of the NH protons parallel to the CH protons of the p-xylene bridge. A further optimization with B3LYP-D3/def2-TZVP gave a ΔE value of 1.04 kcal/mol; however, the ΔG value was only 0.26 kcal/mol. A single-point calculation of these conformations using B2GP-PLYP-D3/def2-TZVPP only slightly changed the ΔE value to 1.08 kcal/mol. Keeping in mind that the typical error range of the used methods is higher than the relative energies found with the DFT methods, no unambiguous preference for one of the tautomers could be given. It thus seemed advisible to change the strategy and perform DLPNO-CCSD(T)/def2-TZVP single-point energy calculations, and in parallel, we started NMR investigations to get a more reliable ratio of the tautomers of 13f.

In the case of both tautomers being equally present for each enantiomer in the racemic mixture, one would expect two NH signals in a 1:1 ratio, since the protons are identical within the molecule but diastereotopic between the two tautomers. In the case, however, that one of the two diasteromeric tautomers should prevail, the two signals should reflect the diastereomeric ratio. Our NMR investigations showed a single signal for the NH protons, which we expected to result from a rapid interconversion of the two tautomers, which is well-known for porphyrins.⁴⁵ NMR investigations at lower temperatures to slow down the interconversion of the tautomers and to detect them side by side⁴⁶ surprisingly did not show any hints at diastereomers appearing, even at temperatures of 225 K. Finally

2D NMR experiments unequivocally proved that already at room temperature only one tautomer was present for each enantiomer (Figure 6). This was also corroborated by high-

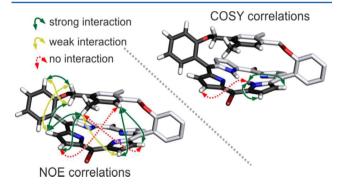


Figure 6. Selected NOE and COSY correlations of the chiral free base BHPs, here shown for S_{p} -**13f** as an example. Selected protons and solubilizing groups are omitted for clarity. Copies of the NOE and COSY spectra of **13f** and **10f**, h are included in the Supporting Information.

level quantum chemical calculations with DLPNO-CCSD(T)/ def2-TZVP, which no longer predicted an equilibrium of the two tautomers but correctly provided an energy difference of $\Delta E = 3.24$ kcal/mol: i.e., large enough to result in only one prevailing tautomer. NOE interactions of the methyl group of the xylene unit to the β -hydrogens of only one pyrrole and of the aromatic hydrogen of the strap to only the other pyrrolic β hydrogens allowed unambiguous differentiation of these pyrrole units. COSY interactions positioned the NH-containing pyrrole unit in proximity to the aromatic hydrogen of the xylene strap, while the other pyrrole clearly was located close to the methyl group of the strap. As expected from the calculations, this combination of NOE and COSY correlations thus established tautomer A as the only existing one, the NH protons being

assigned to the nitrogen atoms orthogonal to the methyl groups of the bridge (Figure 6). This arrested tautomerism is remarkable, since the BHPs are highly symmetric; the tetrapyrrolic framework without the strap is even $C_{2\nu}$ symmetric, which means all pyrrole units would be identical. The loss of the mirror planes resulting in C₂ symmetry is caused only by the unsymmetric substitution of the strap unit, meaning that the energy difference of the two possible tautomers is not due to the presence of an electronically or sterically modified porphyrin macrocycle but is induced solely from the periphery. Furthermore, it is surprising that this phenomenon occurs even at room temperature. Usually temperatures around 200 K or below are necessary to observe arrested tautomerism in porphyrins, even in systems that are electronically unsymmetric: e.g., by β -substitution.⁴⁵ Dispersive interactions, e.g. $\pi - \pi$ stacking, are an explanation of the energy difference of the two tautomers. This can be deduced from the quantum-chemical calculations, which show a distance of 3.40 Å between the planes of the xylene unit and the porphyrin macrocycle for tautomer A, while this distance is 3.50 Å for tautomer B, clearly indicating a stronger interaction between the bridge and the porphyrin for tautomer A.

With the correct ratio of the tautomers in hand, we were quite surprised about the bad matching of the TDDFT calculated spectra with the experimental ECD curves. Even SCS-CC2/def2-SVP results did not fit with the initially measured curves. As mentioned above, we originally tried to acquire ECD spectra of the free base BHPs by online measurements on an HPLC-ECD system, but it seems now that these results were not reliable.³⁸ Therefore, we resolved the enantiomers of 13f by HPLC on a chiral phase on a semipreparative column (Chiralpak IA, n-hexane/dichloromethane 20/80) and measured offline UV/vis (see the Supporting Information) and ECD spectra (Figure 7). The results showed that only within a small window of high dilution $(c = 1 \times 10^{-5} \text{ M})$ was the absorption of the Soret band still within the limits of the Beer-Lambert law, while at the same time it was high enough to permit an ECD signal to be detectable. However, the ECD signals were still not comparable with those of the metalated BHPs, because the free base BHP did not show a couplet in the Soret band region but more or less a single maximum/minimum. The concentration effect described here has been found for all investigated chiral free base BHPs, and thus HPLC-ECD measurements of these have to be done very cautiously or, better, verified with offline ECD spectra.

Finally, only the combination of high-quality experimental measurements (NMR, offline ECD) with high-level calculations (DLPNO-CCSD(T), SCS-CC2) permitted an unambiguous determination of the absolute configurations of the enantiomers of the free base BHP 13f (exemplarily shown for the slower-eluting enantiomer in Figure 7b). These results showed that the enantiomer with a positive peak at ~430 nm did have the R_{p} , P, P configuration, while that with the negative signal was S_{p} , M, M-configured. The arrested tautomerism occurred in all chiral free base BHPs we have synthesized so far, and thus the absolute configurations of these compounds were determined by simple comparison of their experimental ECD spectra with those of 13f (in the case of 10f we again confirmed this determination with SCS-CC2 calculations; see Figure S3 in the Supporting Information).

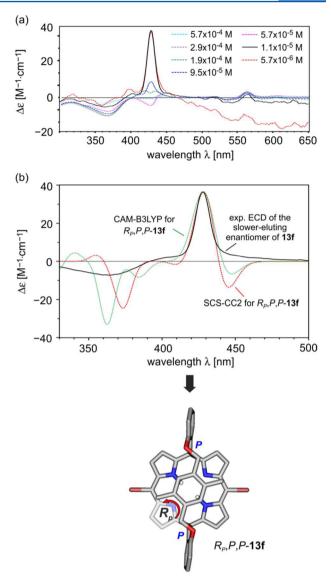


Figure 7. (a) ECD spectra of the slower-eluting enantiomer of 13f measured at different concentrations. (b) Determination of the absolute configuration of the slower-eluting enantiomer of 13f by comparison of its experimental ECD spectrum with those calculated for R_{p} , P, P-13f (the intensities of the calculated spectra are scaled to fit the signal at 430 nm of the experimental curve).

CONCLUSIONS

In summary, we have shown that the BHPs can be modified and functionalized in various positions, which permits the design of a plethora of different structures that can be tuned to their desired purpose. A total of more than 80 different examples are presented in this paper (for a summarizing table see the Supporting Information). It is possible to introduce a broad variety of metals into the porphyrin center, including In(III) or V(IV). The crystal structures of the BHPs shown here gave further insight into this interesting class of compounds and helped to further evaluate the reliability of the ECD calculations. The arrested tautomerism of the chiral free base BHPs induced by the strap is so far unique for porphyrins, and further investigations to utilize this property are under way and will be published in an upcoming paper. In addition, we have successfully resolved several racemates of chiral BHPs to give the pure enantiomers and have elucidated

their absolute configurations by a combination of experimental and computational ECD investigations. We have shown that TD B3LYP suffers from ghost states and that TD CAM-B3LYP should in general be preferred to calculate excited states of BHPs.

COMPUTATIONAL DETAILS

The optimizations and single-point calculations were performed with ORCA.47,48 All functionals were used with dispersion corrections (D3).^{49,50} The chain of spheres⁵¹ approximation was applied for the hybrid and double-hybrid functionals (B3LYP-D3, 52,53 PW6B95-D3, 5 B2GP-PLYP-D3⁵⁵) and RI for B97-D3.⁵⁶ If not stated otherwise, the def2-TZVP⁵⁷ basis set was used. For the tautomers of 13f additionally DLPNO-CCSD(T)58,59 computations have been performed. In all cases, except for V-10d, the tert-butyl or tert-octyl groups have been replaced by hydrogens to save computational time. All TD DFT calculations have been done with Gaussian09⁶⁰ using the functionals B3LYP, CAM-B3LYP, and BHLYP in combination with the def2-SVP⁵⁷ basis set (def2-TZVP for bromine and metal atoms). The SCS-CC2⁶¹ calculations were performed with Turbomole in combination with the def2-SVP basis set (def2-TZVP for metal atoms) and using the RI approximation. Evaluation of the computed and experimental ECD spectra were done with SpecDis^{38,62} applying σ values of 0.08 eV (Ni-10b, Zn-10b, 10f), 0.04 eV (V-10d), or 0.1 eV (13f) and the following UV shifts:⁶³ Ni-10b, 52 nm (SCS-CC2), 48 nm (CAM-B3LYP), 56 nm (BHLYP), 30 nm (B3LYP); Zn-10b, 38 nm (SCS-CC2), 45 nm (CAM-B3LYP), 55 nm (BHLYP), 40 nm (B3LYP); 13f, 68 nm (SCS-CC2), 58 nm (CAM-B3LYP); 10f, 60 nm (SCS-CC2), 53 nm (CAM-B3LYP); V-10d, 55 nm (small gap between strap and porphyrin), 62 nm (large gap) (BHLYP).

EXPERIMENTAL SECTION

General Considerations. All reagents were obtained from commercial sources and used as received. THF was purified and dried by distillation from potassium. All other solvents were used as technical grade. Unless otherwise stated, all reactions were carried out under an atmosphere of dry nitrogen or argon using oven-dried (120 °C) glassware. Analytical TLCs were performed on ready-made plates coated with silica gel on aluminum. Flash chromatography was performed using silica gel 60 (230-400 mesh). ¹H and ¹³C NMR spectra were recorded in CDCl₃ at ambient temperature on spectrometers operating at 400 MHz for ¹H. The ¹³C nucleus was observed with ¹H decoupling. Solvent residual signals were used as an internal standard. Chemical shifts (δ) and coupling constants (J) are given in ppm and Hz, respectively. The peak patterns are indicated as the following format multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; dd, doublet of doublets; dt, doublet of triplets; etc.). HRMS (ESI-TOF) spectra were measured in the positive mode. Analytical enantiomeric resolution was performed on a standard HPLC system equipped with Diacel Chiralpak IA (4.6×250 mm; 3 μ m) as a chiral phase. Online CD spectra were recorded at room temperature (scanning rate 500 nm/min, bandwidth 5 or 10 nm, response time 0.5 s) by HPLC-CD coupling in the stopped-flow mode. Recycling gel permeation chromatography (recycling GPC) was performed on an HPLC system with two columns in line (SDV material, particle size 10 μ m, pore size 50 Å, dimension 20 × 600 mm; SDV material, particle size 10 μ m, pore size 100 Å, dimension 20 \times 600 mm) at a flow rate of 4.5 mL/min in amylene-stabilized chloroform. Salicylic aldehydes $7c^{64}$ and $7d^{65}$ were prepared by reported procedures. Chromatographic details for the enantiomeric resolution of the chiral representatives are included in the Supporting Information. Crystal data collection and processing parameters are given in the Supporting Information. The BHPs in the Experimental Section are denoted by their compound number, and brief information regarding their substitution is added ("strap: 2Me/4Me/2Br/4H" describes the substitution R¹/R² of the xylene unit, "H-/t-Bu-/t-Octaryl" describes the presence of substituents R⁵ (solubilizing groups) at

the meso-aryls, "meso:" denotes the substituents at the meso-positions R^3/R^4 not occupied by the basket-handle strap).

General Procedure A: Preparation of the Strap-Dialdehydes 9. The dibromoxylene-strap 6 (20 mmol, 1.0 equiv), salicylaldehyde 7 (42 mmol, 2.1 equiv), powdered anhydrous K_2CO_3 (14.0 g, 0,10 mol, 5.0 equiv), and 600 mL of acetone were heated to reflux for 10 h. The solvent was removed, and the remaining solid was suspended in 400 mL of chloroform. Dilute HCl was added until all material was dissolved. The organic phase was separated, washed with NaHCO₃ solution, and dried over NaSO₄. Evaporation of the solvent yielded **9** in sufficient purity for the condensation step. An analytical sample was recrystallized from ethyl acetate to afford white flakes. Strap-dialdehydes **9a**–c were reported before.¹²

6,6'-((2,5-Dimethyl-1,4-phenylene)dimethylenedioxy)bis(3-(tertbutyl))dibenzaldehyde (9d). Prepared from dibromoxylene-strap 6d (5.84 g) and salicylaldehyde 7b (7.48 g). White solid, 9.43 g, 97% yield; mp 146−150 °C. ¹H NMR (400 MHz, CDCl₃): δ 1.32 (s, 18H), 2.37 (s, 6H), 5.13 (s, 4H), 7.04 (d, ³J = 8.8 Hz, 2H), 7.29 (s, 2H), 7.61 (dd, ³J = 8.7 Hz, ⁴J = 2.7 Hz, 2H), 7.89 (d, ⁴J = 2.6 Hz, 2H), 10.52 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 190.3, 159.6, 144.3, 134.6, 134.5, 133.5, 131.0, 125.3, 124.8, 113.0, 77.7, 77.6, 77.4, 77.0, 69.2, 34.6, 31.6, 18.9 ppm. MS (EI): *m*/*z* 309.1 (60) [M − salicylic aldehyde group]^{•+}, 132.18 (100) [M − 2 × salicylic aldehyde group]^{•+}, 102.0 (70). HRMS (ESI): calcd for C₃₂H₃₈NaO₄ [M + Na]⁺ 509.26623, found 509.26611.

6,6'-((2,3,5,6-Tetramethyl-1,4-phenylene)dimethylenedioxy)bis-(3-(tert-octyl))dibenzaldehyde (**9e**). Prepared from dibromoxylenestrap **6b** (6.40 g) and salicylaldehyde 7c (9.83 g). White solid, 11.6 g, 97% yield; mp 208–210 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.75 (s, 18H), 1.39 (s, 12H), 1.76 (s, 4H), 2.36 (s, 12H), 5.22 (s, 4H), 7.16 (d, ³J = 8.8 Hz, 2H), 7.64 (dd, ³J = 8.8 Hz, ⁴J = 2.7 Hz, 2H), 7.88 (d, ⁴J = 2.7 Hz, 2H), 10.40 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 16.5, 31.7, 32.0, 32.5, 38.3, 56.8, 66.3, 112.5, 124.6, 125.6, 133.1, 134.0, 134.9, 143.2, 159.7, 190.2 ppm. MS (EI): *m/z* 393.4 (100) [M – salicylic aldehyde group]^{•+}, 160.2 (99) [M – 2 × salicylic aldehyde group]^{•+}, 57.1 (22). HRMS (ESI): calcd for C₄₂H₃₈NaO₄ [M + Na]⁺ 649.42273, found 649.42253.

6,6'-((2,5-Dimethyl-1,4-phenylene)dimethylenedioxy)bis(3-(tert-octyl))dibenzaldehyde (9f). Prepared from dibromoxylene-strap 6d (5.84 g) and salicylaldehyde 7c (9.83 g). White solid, 12.3 g, 98% yield; mp 155–159 °C. ¹H NMR (400 MHz, CCl₃): δ 0.72 (s, 18H), 1.37 (s, 12H), 1.74 (s, 4H), 2.36 (s, 6H), 5.13 (s, 4H), 7.02 (d, ³J = 8.7 Hz, 2H), 7.28 (s, 2H), 7.58 (dd, ³J = 8.7 Hz, ⁴J = 2.7 Hz, 2H), 7.87 (d, ³J = 2.7 Hz, 2H), 10.52 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 18.7, 31.6, 32.0, 32.5, 38.3, 56.9, 69.0, 77.4, 112.6, 124.5, 125.8, 130.8, 134.1, 134.3, 134.4, 143.2, 159.3, 190.1 ppm. MS (EI): *m*/*z* 365.4 (100) [M – salicylic aldehyde group]⁺⁺, 132.2 (91) [M – 2 × salicylic aldehyde group]⁺⁺, 57.1 (20). HRMS (ESI): calcd for C₄₀H₅₄NaO₄ [M + Na]⁺ 621.39143, found 621.39253.

6, 6' - ((2, 5-Dibromo-1, 4-phenylene)dimethylenedioxy)dibenzaldehyde (**9g**). Prepared from dibromoxylene-strap **6c** (8.44 g) and salicylaldehyde **7a** (5.12 g). White solid, 9.54 g, 95% yield; mp 203–207 °C. ¹H NMR (400 MHz, CDCl₃): δ 5.23 (s, 4H), 7.05 (m, 2H), 7.12 (m, 2H), 7.59 (m, 2H), 7.83 (s, 2H), 7.90 (m, 2H), 10.59 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 189.7, 160.5, 137.4, 136.4, 132.9, 129.5, 125.6, 122.2, 122.1, 121.6, 113.3, 77.7, 77.6, 77.4, 77.0, 69.5 ppm. MS (EI): *m*/*z* 381.8 (62) [M – salicylic aldehyde group]^{•+}, 261.8 (100) [M – salicylic aldehyde group, –CHO]^{•+}, 102.0 (70). HRMS (ESI): calcd for C₂₂H₁₆Br₂NaO₄ [M + Na]⁺ 524.93076, found 524.93038.

6,6'-((2,5-Dibromo-1,4-phenylene)dimethylenedioxy)bis(3-(tert-octyl))dibenzaldehyde (9h). Prepared from dibromoxylene-strap 6c (8.44 g) and salicylaldehyde 7c (9.83 g). White solid, 13.8 g, 98% yield; mp 77–80 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.72 (s, 18H), 1.38 (s, 12H), 1.74 (s, 4H), 5.21 (s, 4H), 6.97 (d, ³J = 8.8 Hz, 2H), 7.59 (dd, ³J = 8.8 Hz, ⁴J = 2.6 Hz, 2H), 7.82 (s, 2H), 7.90 (d, ⁴J = 2.6 Hz, 2H), 10.59 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 209.8, 152.2, 143.8, 138.0, 132.8, 130.5, 126.2, 124.9, 121.5, 111.2, 69.1, 66.0, 57.3, 51.0, 38.6, 32.7, 32.2, 32.1, 32.1, 31.8, 31.2 ppm. MS (EI): *m/z* 495.2 (82) [M – salicylic aldehyde group]*, 423.1 (53), 262.0 (93)

 $[M - 2 \times salicylic aldehyde group]^{\bullet+}$, 161.1 (100), 57.1 (92). HRMS (ESI): calcd for $C_{38}H_{48}Br_2O_4$ $[M]^+$ 726.19194, found 726.19209.

6,6'-((1,4-Phenovlene)dimethylenedioxy)bis(3-(tert-octyl))dibenzaldehyde (9i). Prepared from dibromoxylene-strap 6a (5.28 g) and salicylaldehyde 7c (9.83 g). White solid, 11.0 g, 96% yield; mp 111–115 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.71 (s, 18H), 1.36 (s, 12H), 1.73 (s, 4H), 5.19 (s, 4H), 6.98 (d, ³J = 8.8 Hz, 2H), 7.48 (s, 4H), 7.56 (dd, ³J = 8.8 Hz, ⁴J = 2.7 Hz, 2H), 7.86 (d, ³J = 2.6 Hz, 2H), 10.55 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 190.3, 159.2, 143.4, 136.7, 134.2, 128.0, 126.1, 124.6, 112.8, 77.7, 77.6, 77.4, 77.0, 70.5, 57.0, 38.5, 32.7, 32.2, 32.1, 31.8 ppm. MS (EI): m/z 337.3 (76) [M – salicylic aldehyde group]^{•+}, 57.1 (40). HRMS (ESI): calcd for C₃₈H₅₀NaO₄ [M + Na]⁺ 593.36013, found 593.36065.

6,6'-((2,3,5,6-Tetramethyl-1,4-phenylene)dimethylenedioxy)bis-(3-phenyl)dibenzaldehyde (9j). Prepared from dibromoxylene-strap 6b (6.40 g) and salicylaldehyde 7d (8.14 g). White solid, 10.8 g, 97% yield; mp 211–216 °C. ¹H NMR (400 MHz, CDCl₃): δ 2.39 (s, 12H), 5.30 (s, 4H), 7.33 (d, ³J = 8.6 Hz, 2H), 7.37 (d, ³J = 7.4 Hz, 2H), 7.44–7.47 (m, 4H), 7.61 (m, 4H), 7.87, (dd, ³J = 8.6 Hz, ⁴J = 2.5 Hz, 2H), 8.13 (d, ⁴J = 2.5 Hz, 2H), 10.46 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 16.6, 66.4, 113.4, 125.5, 126.8, 126.9, 127.5, 133.0, 134.4, 134.5, 135.0, 139.7, 161.1, 189.9 ppm. MS (EI): m/z 357.3 (76) [M – salicylic aldehyde group]^{•+}, 160.1 (100) [M – 2 × salicylic aldehyde group]^{•+}. HRMS (ESI): calcd for C₃₈H₃₄NaO₄ [M + Na]⁺ 577.23493, found 577.23386.

6,6'-((2,5-Dibromo-1,4-phenylene)dimethylenedioxy)bis(3phenyl)dibenzaldehyde (9k). Prepared from dibromoxylene-strap 6d (5.84 g) and salicylaldehyde 7d (8.14 g). White solid, 10.2 g, 95% yield; mp 180–186 °C. ¹H NMR (400 MHz, CDCl₃): δ 1.26 (s, 2H), 1.54 (s, 15H), 2.41 (s, 6H), 5.21 (s, 4H), 7.19 (d, ³J = 8.7 Hz, 2H), 7.33–7.37 (m, 4H), 7.43–7.46 (m, 4H), 7.57–7.61 (m, 54H), 7.82 (dd, ³J = 8.6 Hz, ⁴J = 2.5 Hz, 2H), 8.12 (d, ⁴J = 2.5 Hz, 2H), 10.58 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 18.6, 69.1, 113.4, 125.3, 126.7, 126.8, 127.4, 128.9, 130.8, 134.2, 134.3, 134.3, 139.5, 160.6, 189.6 ppm. MS (EI): m/z 329.2 (100) [M – salicylic aldehyde group]^{•+}, 197 (45), 132.1 (94) [M – 2 × salicylic aldehyde group]^{•+}. HRMS (ESI): calcd for C₃₆H₃₀NaO₄ [M + Na]⁺ 549.20363, found 549.20486.

General Procedure B: Condensation of Strapped Dialdehydes with Dipyrromethane. Dipyrromethane 8a (2.92 g, 0.02 mol, 2.0 equiv) and the strap-dialdehyde 9 (0.01 mol, 1 equiv) were dissolved in dichloromethane (900 mL), and the solution was degassed by an argon stream for 20 min. The reaction mixture was shielded from ambient light, TFA (1.5 mL) was added, and the mixture was stirred at room temperature until TLC indicated full consumption of the starting material (between 3 and 5 h). NEt₃ (5 mL) and p-chloranil (14.7 g, 0.06 mol, 6 equiv) were added, and the mixture was heated to reflux for 1 h. After evaporation of about 700 mL of dichloromethane, n-hexane (300 mL) was added and the black solution was filtered over a plug of silica (150 g) and washed with dichloromethane/n-hexane 2/1, which yielded a purple solution. After evaporation of the solvents, the porphyrin was recrystallized from chloroform/methanol to yield purple crystals of high purity. BHPs 10a-c were reported before.

BHP 10d (strap: 2Me, *t*-Bu-aryl, *meso*: 2H). Prepared from strapdialdehyde **9d** (4.86 g). 2.45 g, 30% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –3.41 (s, 2H), –0.66 (s, 6H), 1.77 (s, 18H), 2.23 (d, ³*J* = 13.2 Hz, 2H), 3.39 (app t, ^{app}*J* = 6.6 Hz 4H), 7.01 (d, ³*J* = 8.4 Hz, 2H), 7.68 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.6 Hz, 2H), 8.83 (d, ³*J* = 4.6 Hz, 2H), 9.06 (d, ³*J* = 4.5 Hz, 2H), 9.09 (d, ⁴*J* = 2.6 Hz, 2H), 9.21 (d, ³*J* = 4.5 Hz, 2H), 9.23 (d, ³*J* = 4.6 Hz, 2H), 9.97 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.5, 32.2, 35.0, 72.3, 104.3, 112.5, 120.6, 120.9, 126.2, 126.2, 126.7, 128.3, 129.9, 130.4, 130.5, 132.1, 133.7, 134.1, 145.9, 158.9 ppm. MS (MALDI): *m*/*z* 736.312 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₉N₄O₂ [M + H]⁺ 737.38555, found 737.38500.

BHP 10e (strap: 4Me, *t*-Oct-aryl, *meso*: 2H). Prepared from strapdialdehyde **9e** (6.26 g). 2.80 g, 32% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.91 (s, 2H), -0.05 (s, 12H), 1.17 (s, 18H), 1.81 (s, 12H), 2.17 (s, 4H), 3.61 (s, 4H), 6.56 (d, ³J = 8.3 Hz, 2H), 7.58 (dd, ${}^{3}J$ = 8.4 Hz, ${}^{4}J$ = 2.5 Hz, 2H), 8.83 (d, ${}^{3}J$ = 4.6 Hz, 4H), 9.04 (d, ${}^{4}J$ = 2.4 Hz, 2H), 9.13 (d, ${}^{3}J$ = 4.6 Hz, 4H), 9.92 (s, 2H) ppm. 13 C NMR (100 MHz, CDCl₃): δ 13.4, 29.9, 32.5, 32.5, 33.0, 38.7, 57.5, 63.6, 103.8, 110.0, 114.0, 126.4, 127.2, 130.5, 130.5, 130.9, 140.9, 156.3 ppm. MS (MALDI): m/z 876.492 [M]*+. HRMS (ESI): calcd for C₆₀H₆₉N₄O₂ [M + H]⁺ 877.54150, found 877.54208.

BHP 10f (strap: 2Me, *t*-Oct-aryl, *meso*: 2H). Prepared from strapdialdehyde **9f** (5.98 g). 2.54 g, 30% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -3.40 (s, 2H), -0.64 (s, 6H), 1.17 (s, 18H), 1.81 (s, 6H), 1.84 (s, 6H), 2.14 (d, ³*J* = 14.6 Hz, 2H), 2.21 (d, ³*J* = 14.6 Hz, 2H), 2.25 (d, ³*J* = 13.5 Hz, 2H), 3.40 (s, 2H), 3.43 (s, 1H), 7.02 (d, ³*J* = 8.4 Hz, 2H), 7.69 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 8.81 (d, ³*J* = 4.6 Hz, 2H), 9.05 (d, ³*J* = 4.5 Hz, 2H), 9.09 (d, ⁴*J* = 2.5 Hz, 2H), 9.21 (d, ³*J* = 4.5 Hz, 2H), 9.24 (d, ³*J* = 4.6 Hz, 2H), 9.97 (s, 2H) ppm. ¹³C NMR (150 MHz, CDCl₃): δ 14.5, 32.0, 32.5, 32.9, 33.0, 38.9, 57.5, 72.3, 104.3, 112.6, 120.4, 120.9, 126.2, 127.7, 129.1, 130.4, 133.9, 144.7, 158.8 ppm. MS (MALDI): *m*/*z* 848.675 [M]^{•+}. HRMS (ESI): calcd for C₅₈H₆₅N₄O₂ [M + H]⁺ 849.51020, found 849.51107.

BHP 10g (strap: 2Br, H-aryl, *meso*: 2H). Prepared from strapdialdehyde 9g (5.02 g). 1.96 g, 26% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –3.38 (s, 2H), 2.16 (d, ³J = 14.8 Hz, 2H), 3.33 (d, ³J = 14.8 Hz, 2H), 3.78 (s, 2H), 7.11 (dd, ³J = 7.9 Hz, ⁴J = 1.2 Hz, 2H), 7.69 (td, ³J = 7.9 Hz, ⁴J = 1.8 Hz, 2H), 7.81 (td, ³J = 7.9 Hz, ⁴J = 1.3 Hz, 2H), 8.76 (d, ³J = 4.6 Hz, 2H), 9.04 (dd, ³J = 7.4 Hz, ⁴J = 1.8 Hz, 2H), 9.07 (d, ³J = 4.6 Hz, 2H), 9.25 (dd, ³J = 4.6 Hz, ⁴J = 1.2, 4H), 10.04 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 160.7, 135.1, 133.3, 131.6, 130.4, 125.1, 123.9, 122.2, 114.6, 111.1, 105.4, 77.7, 77.6, 77.4, 77.0, 73.5 ppm. MS (MALDI): *m*/z 751.957 [M]^{•+}. HRMS (ESI): calcd for C₄₀H₂₇Br₂N₄O₂ [M + H]⁺: 753.04953, found 753.05002.

BHP 10h (strap: 2Br, *t*-Oct-aryl, *meso*: 2H). Prepared from strapdialdehyde **9h** (7.04 g). 3.12 g, 32% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -3.33 (s, 2H), 1.17 (s, 18H), 1.82 (d, ³*J* = 13.1 Hz, 12H), 2.12–2.23 (m, 6H), 3.33 (d, ³*J* = 14.6 Hz, 2H), 3.80 (s, 2H), 7.03 (d, ³*J* = 8.4 Hz, 2H), 7.70 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.4 Hz, 2H), 8.76 (d, ³*J* = 4.6 Hz, 2H), 9.05 (d, ³*J* = 4.6 Hz, 2H), 9.10 (d, ⁴*J* = 2.4 Hz, 2H), 9.24–9.26 (m, 4H), 10.04 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 31.9, 32.5, 32.9, 33.0, 38.9, 57.5, 73.3, 105.1, 111.6, 114.4, 121.0, 124.9, 127.7, 129.4, 133.2, 133.9, 145.4, 158.3 ppm. MS (MALDI): *m/z* 976.316 [M]^{•+}. HRMS (ESI): calcd for C₅₆H₅₉Br₂N₄O₂ [M + H]⁺ 977.29993, found 977.30022.

BHP 10i (strap: 4H, *t*-Oct-aryl, *meso*: 2H). Prepared from strapdialdehyde 9i (5.71 g). 1.97 g, 24% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.51 (s, 2H), 1.14 (s, 18H), 1.81 (s, 12H), 2.16 (s, 4H), 3.08 (s, 4H), 3.32 (s, 4H), 6.73 (d, ³*J* = 8.4 Hz, 2H), 7.62 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 8.85 (d, ³*J* = 4.5 Hz, 4H), 9.10 (d, ⁴*J* = 2.4 Hz, 2H), 9.20 (d, ³*J* = 4.6 Hz, 4H), 9.98 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 157.3, 143.1, 133.9, 132.4, 132.2, 131.2, 128.0, 127.8, 123.1, 115.5, 113.6, 104.7, 71.3, 57.7, 38.9, 33.2, 32.7, 32.6, 30.1 ppm. MS (MALDI): *m*/*z* 820.445 [M]^{•+}. HRMS (ESI): calcd for C₅₆H₆₁N₄O₂ [M + H]⁺ 821.47890, found 821.48016.

General Procedure C: meso-Substituted BHPs via Mixed Condensation of Strap-Dialdehydes with Dipyrromethanes. meso-unsubstituted dipyrromethane 8a⁶⁶ (1.46 g, 0.01 mol, 1.0 equiv), meso-substituted dipyrromethane 8⁶⁶ (0.01 mol, 1.0 equiv), and the strap-dialdehyde 9 (0.01 mol, 1 equiv) were dissolved in dichloromethane (900 mL), and the solution was degassed by an argon stream for 20 min. The reaction mixture was shielded from ambient light, TFA (1.5 mL) was added, and the mixture was stirred at room temperature until TLC indicated full consumption of the starting material (between 3 and 5 h). NEt₃ (5 mL) and p-chloranil (14.7 g, 0.06 mol, 6 equiv) were added, and the mixture was heated to reflux for 1 h. After evaporation of about 700 mL of dichloromethane, nhexane (300 mL) was added and the black solution was filtered over a plug of silica (150 g) and washed with dichloromethane/*n*-hexane 2/1, which yielded a purple solution. Column chromatography (silica, dichloromethane/n-hexane) yielded meso-substituted BHP 11 along with meso-unsubstituted BHP 10 and meso-disubstituted BHP 12. Analytical samples were recrystallized from chloroform/methanol to afford purple crystals.

BHP 11a/12a (strap: 4Me, H-aryl; meso: (4-t-Bu)Ph). Prepared from substituted dipyrromethane 8b (2.78 g) and strap-dialdehyde 9a (4.02 g), column chromatography with DCM/n-hexane 30/70. 10a (strap: 4Me, H-aryl; meso: $2 \times H$): 0.45 g, 7%, characterization identical with that reported before. 11a (strap: 4Me, H-aryl; meso: H/ (4-t-Bu)Ph): 1.09 g, 14% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.57 (s, 2H), 0.05 (s, 6H), 0.13 (s, 6H), 1.60 (s, 9H), 3.71 (s, 4H), 6.68 (dd, ³J = 7.9 Hz, ⁴J = 1.4 Hz, 4H), 7.54–7.62 (m, 4H), 7.74 (d, ${}^{3}J$ = 8.4 Hz, 2H), 8.06 (s, 2H), 8.73–8.76 (m, 4H), 8.84 (d, ${}^{3}J$ = 4.6 Hz, 2H), 8.95 (dd, ${}^{3}J$ = 7.0 Hz, ${}^{4}J$ = 1.9 Hz, 2H), 9.08 (d, ${}^{3}J$ = 4.6 Hz, 2H), 9.79 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.5, 13.5, 29.9, 31.8, 35.5, 63.6, 102.9, 110.8, 113.7, 119.4, 120.1, 123.8, 128.1, 130.0, 130.5, 131.0, 131.1, 131.2, 131.6, 134.2, 139.1, 150.4, 158.7 ppm. MS (MALDI): *m*/*z* 784.319 [M]^{•+}. HRMS (ESI): calcd for $C_{54}H_{49}N_4O_2$ $[M + H]^+$ 785.38500, found 785.38671. 12a (strap: 4Me, H-aryl; meso: $2 \times (4-t-Bu)Ph$): 0.55 g, 6% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.19 (s, 2H), 0.18 (s, 12H), 1.60 (s, 18H), 3.76 (s, 4H), 6.69 (dd, ${}^{3}J$ = 7.8 Hz, ${}^{4}J$ = 1.3 Hz, 2H), 7.49–7.61 (m, 4H), 7.74 (d, ${}^{3}J$ = 8.6 Hz, 4H), 8.06 (d, ${}^{3}J$ = 7.7 Hz, 4H), 8.72 (d, ${}^{3}J = 4.7$ Hz, 4H), 8.75 (d, ${}^{3}J = 4.8$ Hz, 4H), 8.92 (dd, ${}^{3}J = 7.1$ Hz, ${}^{4}J =$ 1.9 Hz, 2H) ppm. ¹³C NMR (150 MHz, CDCl₃): δ 13.6, 16.1, 31.6, 31.8, 35.0, 63.8, 77.4, 110.9, 114.0, 119.1, 119.4, 123.9, 125.8, 127.9, 128.2, 129.9, 130.7, 130.9, 131.0, 131.1, 131.9, 134.3, 138.9, 140.8, 146.0, 146.4, 150.4, 158.8 ppm. MS (MALDI): m/z 916.438 [M]^{•+}. HRMS (ESI): calcd for C₆₄H₆₁N₄O₂ [M + H]⁺ 917.47890, found 917.48068

BHP 11b/12b (strap: 4Me, t-Bu-aryl; meso: (4-t-Bu)Ph). Prepared from substituted dipyrromethane 8b (2.78 g) and strap-dialdehyde 9c (5.14 g), column chromatography with DCM/*n*-hexane $30/70 \rightarrow 50/$ 50. 10c (strap: 4Me, t-Bu-aryl; meso: $2 \times H$): 0.38 g, 5% yield; characterization identical with that reported before. 11b (strap: 4Me, t-Bu-aryl; meso: H/(4-t-Bu)Ph): 1.34 g, 15% yield; mp >300 °C. ¹H NMR (400 MHz, $CDCl_3$): δ -2.51 (s, 2H), 0.06 (s, 6H), 0.13 (s, 6H), 1.63 (s, 18H), 3.71 (s, 4H), 6.62 (d, ${}^{3}J$ = 8.5 Hz, 2H), 7.62 (dd, ${}^{3}J$ = 8.5 Hz, ⁴J = 2.6 Hz, 2H), 7.76 (d, ³J = 8.5 Hz, 2H), 8.09 (s, 2H), 8.76-8.79 (m, 4H), 8.86 (d, ³*J* = 4.7 Hz, 2H), 9.05 (d, ⁴*J* = 2.6 Hz, 2H), 9.07 (d, ${}^{3}J$ = 4.7 Hz, 2H), 9.78 (s, 1H) ppm. ${}^{13}C$ NMR (100 MHz, CDCl₂): δ 13.5, 13.6, 31.8, 32.2, 34.8, 35.0, 63.9, 102.8, 110.3, 114.2, 120.1, 123.8, 125.7, 126.1, 130.7, 130.9, 131.1, 131.1, 131.3, 134.3, 139.1, 142.2, 150.4, 156.5 ppm. MS (MALDI): m/z 896.478 [M]^{•+}. HRMS (ESI): calcd for $C_{62}H_{65}N_4O_2$ [M + H]⁺ 897.51020, found 897.50939. 12b (strap: 4Me, t-Bu-aryl; meso: 2 × (4-t-Bu)Ph): 0.72 g, 7% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.13 (s, 2H), 0.19 (s, 12H), 1.63 (s, 18H), 1.77 (s, 18H), 3.77 (s, 4H), 6.64 (d, ³J = 8.5 Hz, 2H), 7.61 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 7.77 (d, ³*J* = 8.5 Hz, 4H), 8.10 (d, ${}^{3}J$ = 7.5 Hz, 4H), 8.76 (d, ${}^{3}J$ = 4.7 Hz, 2H), 8.78 (d, ${}^{3}J$ = 4.7 Hz, 2H), 9.03 (d, ${}^{4}J$ = 2.4 Hz, 2H) ppm. 13 C NMR (100 MHz, CDCl₃): *δ* 13.6, 31.8, 32.2, 34.7, 35.0, 64.1, 110.5, 114.6, 119.0, 123.9, 125.8, 126.1, 130.8, 130.9, 131.0, 131.4, 134.4, 138.9, 142.2, 145.9, 146.3, 150.3, 156.6 ppm. MS (MALDI): *m*/*z* 1029.604 [M]^{•+}. HRMS (ESI): calcd for $C_{72}H_{77}N_4O_2[M + H]^+$ 1029.6047, found 1029.6002.

BHP 11c/12c (strap: 4Me, t-Bu-aryl; meso: (3,5-di-t-Bu)Ph). Prepared from substituted dipyrromethane 8c (3.34 g) and strapdialdehyde 9c (5.14 g), column chromatography with DCM/n-hexane $20/80 \rightarrow 50/50$. 10c (strap: 4Me, t-Bu-aryl; meso: 2 × H): 0.38 g, 5% yield; characterization identical with that reported before. 11c (strap: 4Me, t-Bu-aryl; meso: H/(3,5-di-t-Bu)Ph): 1.33 g, 14% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.47 (s, 2H), 0.04 (s, 6H), 0.26 (s, 6H), 1.59 (s, 18H), 1.82 (s, 18H), 3.73 (q, 4H), 6.64 (d, ${}^{3}J$ = 8.4 Hz, 2H), 7.65 (d, ${}^{3}J$ = 8.4 Hz, ${}^{4}J$ = 2.5 Hz, 2H), 7.85 (app t, ${}^{app}J$ = 1.8 Hz, 1H), 8.78 (d, ${}^{3}J$ = 4.7 Hz, 2H), 8.82 (d, ${}^{3}J$ = 4.7 Hz, 2H), 8.90 (d, ${}^{3}J$ = 4.6 Hz, 2H), 9.09 (s, 2H), 9.10 (d, ${}^{4}J$ = 1.9 Hz, 2H), 9.79 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.4, 13.5, 31.9, 32.3, 34.8, 35.1, 63.8, 102.8, 110.2, 114.2, 120.9, 121.3, 125.6, 126.1, 129.6, 130.7, 130.7, 130.9, 131.0, 131.2, 131.3, 141.2, 142.2, 148.8, 156.6 ppm. MS (MALDI): m/z 952.425 [M]^{•+}. HRMS (ESI): calcd for C₆₆H₇₃N₄O₂ $[M + H]^+$ 953.57280, found 953.57183. **12c** (strap: 4Me, *t*-Bu-aryl; *meso*: $2 \times (3,5\text{-di-}t\text{-Bu})\text{Ph})$: 0.68 g, 6% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.19 (s, 2H), 0.20 (s, 12H), 1.53 (s, 36H), 1.74 (s, 18H), 3.75 (s, 4H), 6.62 (d, ${}^{3}J$ = 8.4 Hz, 2H), 7.59 (dd, ${}^{3}J$ =

8.4 Hz, ${}^{4}J$ = 2.5 Hz, 2H), 7.78 (app t, ${}^{\rm app}J$ = 1.8 Hz, 2H), 8.71 (d, ${}^{3}J$ = 4.7 Hz, 4H), 8.76 (d, ${}^{3}J$ = 4.7 Hz, 4H), 9.00 (d, ${}^{4}J$ = 1.4 Hz, 2H) ppm. 13 C NMR (100 MHz, CDCl₃): δ 13.5, 31.9, 32.2, 34.7, 35.1, 64.0, 110.3, 114.5, 120.2, 121.0, 125.6, 126.1, 129.6, 130.8, 131.0, 131.0, 131.6, 141.0, 142.2, 146.3, 146.6, 149.0, 156.6 ppm. MS (MALDI): m/z 1140.964 [M]*+. HRMS (ESI): calcd for C₈₀H₉₃N₄O₂ [M + H]+ 1141.72930, found 1141.73052.

BHP 11d/12d (strap: 4Me, t-Oct-aryl; meso: (3,5-di-t-Bu)Ph). Prepared from substituted dipyrromethane 8c (3.34 g) and strapdialdehyde 9e (5.98 g), column chromatography with DCM/n-hexane $20/70 \rightarrow 40/60$. 10e (strap: 4Me, t-Oct-aryl; meso: 2 × H): 0.78 g, 9% yield; characterization identical with that reported above. 11d (strap: 4Me, t-Oct-aryl; meso: H/(3,5-di-t-Bu)Ph): 1.59 g, 15% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.56 (s, 2H), -0.01 (s, 6H), 0.22 (s, 6H), 1.41 (s, 18H), 1.79 (d, ${}^{3}J = 7.9$ Hz, 12H), 2.13 (d, ${}^{3}J$ = 14.7 Hz, 2H), 2.18 (d, ${}^{3}J$ = 14.7 Hz, 2H), 3.67 (d, ${}^{3}J$ = 10.0 Hz, 2H), 3.71 (d, ${}^{3}I = 10.0$ Hz, 2H), 6.59 (d, ${}^{3}I = 8.4$ Hz, 2H), 7.57 (dd, ${}^{3}I = 8.4$ Hz, ${}^{4}J = 2.6$ Hz, 2H), 7.76 (app t, ${}^{app}J = 1.8$ Hz, 2H), 8.70 (d, ${}^{3}J = 4.7$ Hz, 2H), 8.72 (d, ${}^{3}J$ = 4.7 Hz, 2H), 8.83 (d, ${}^{3}J$ = 4.6 Hz, 2H), 9.00 (d, ${}^{4}I = 2.6$ Hz, 2H), 9.09 (d, ${}^{3}I = 4.6$ Hz, 2H), 9.80 (s, 1H) ppm. ${}^{13}C$ NMR (100 MHz, CDCl₃): δ 13.5, 13.5, 31.9, 32.3, 32.5, 32.7, 33.0, 35.1, 38.6, 53.6, 57.5, 63.8, 102.7, 110.0, 114.3, 121.0, 121.2, 126.5, 127.1, 129.3, 130.7, 130.9, 131.0, 131.0, 140.9, 141.2, 156.4 ppm. MS (MALDI): m/z 1064.650 [M]^{•+}. HRMS (ESI): calcd for $C_{74}H_{89}N_4O_2$ [M + H]⁺ 1065.69800, found 1065.69961. **12d** (strap: 4Me, *t*-Oct-aryl; meso: $2 \times (3,5\text{-di-}t\text{-Bu})\text{Ph})$: no tetraaryl-BHP fraction was observed.

General Procedure D: meso-Substitution of BHPs with Organolithium Reagents. The BHP (100 μ mol, 1 equiv) was dissolved in absolute THF (25 mL) and cooled to -10 °C. R-Li (300 μ mol, 3 equiv) was added dropwise over 5 min and the reaction mixture was stirred until no remaining BHP was detected by TLC. Water (2 mL) was added, and the solution was warmed to room temperature. DDQ (91 mg, 400 μ mol, 4 equiv) was added and the mixture was stirred for 1 h. THF was completely removed, and dichloromethane (100 mL) was added. The organic solution was washed with dilute HCl, NaHCO₃, and water and dried over NaSO₄. After evaporation of the solvent the remaining solid was purified by column chromatography (silica, dichloromethane/*n*-hexane). The raw product was recrystallized from chloroform/methanol to yield purple crystals. BHPs 11e,f were reported before.

BHP 11g (strap: 4Me, H-aryl; *meso*: H/*n*-Hex). Prepared from BHP **10a** (65 mg) and *n*-HexLi (0.12 mL, 2.5 M in *n*-hexane), column chromatography with DCM/*n*-hexane 20/70 \rightarrow 50/50. 55 mg, 75% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.41 (s, 2H), 0.04 (s, 6H), 0.13 (s, 6H), 1.35-1.41 (m, 2H), 1.47-1.51 (m, 4H), 1.73-1.77 (m, 2H), 2.46-2.50 (m, 2H), 3.71 (d, ³J = 9.9 Hz, 2H), 3.76 (d, ³J = 9.9 Hz, 2H), 4.82 (app t, ^{app}J = 8.0 Hz, 2H), 6.69 (dd, ³J = 7.9 Hz, ⁴J = 1.5 Hz, 2H), 7.54-7.63 (m, 4H), 8.79 (d, ³J = 4.6 Hz, 2H), 8.82 (d, ³J = 4.6 Hz, 2H), 8.94 (dd, ³J = 7.1 Hz, ⁴J = 1.9 Hz, 2H), 9.02 (d, ³J = 4.6 Hz, 2H), 9.27 (d, ³J = 4.6 Hz, 2H), 9.68 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.5, 14.3, 22.9, 30.3, 32.1, 35.1, 39.0, 63.6, 102.2, 110.8, 113.2, 119.4, 120.7, 128.1, 128.3, 129.9, 130.6, 130.9, 131.0, 131.2, 131.3, 131.6, 158.7 ppm. MS (MALDI): *m*/z 736.298 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₈N₄O₂ [M]⁺ 736.37718, found 736.37766.

BHP 11a (strap: 4Me, H-aryl; *meso*: H/(4-t-Bu)Ph). Prepared from BHP 10a (65 mg) and (4-t-Bu)PhLi (0.1 mL, ~ 3 M in diethyl ether),⁶⁷ column chromatography with DCM/*n*-hexane 20/70 $\rightarrow 50/50$. 51 mg, 65% yield; characterization identical with that reported above.

BHP 11b (strap: 4Me, *t*-Bu-aryl; *meso*: H/(4-*t*-Bu)Ph). Prepared from BHP **10c** (76 mg) and (4-*t*-Bu)PhLi (0.1 mL, ~ 3 M in diethyl ether),⁶⁷ column chromatography with DCM/*n*-hexane $30/70 \rightarrow 50/50$. 61 mg, 68% yield; characterization identical with that reported above.

BHP 11h (strap: 2Me, H-aryl; *meso*: H/*n*-Bu). Prepared from BHP **10b** (63 mg) and *n*-BuLi (0.12 mL, 2.5 M in *n*-hexane), column chromatography with DCM/*n*-hexane 20/70 \rightarrow 50/50. 49 mg, 72% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.95 (s, 2H), -0.61 (s, 3H), -0.53 (s, 3H), 2.44-2.56 (m, 4H), 3.50 (d, ³J = 12.9

Hz, 2H), 3.60 (d, ${}^{3}J$ = 15.9 Hz, 2H), 7.10 (d, ${}^{3}J$ = 7.9 Hz, 2H), 7.66– 7.70 (m, 2H), 7.74–7.78 (m, 2H), 8.73 (dd, ${}^{3}J$ = 12.0 Hz, ${}^{4}J$ = 4.7 Hz, 2H), 8.98–9.00 (m, 4H), 9.03 (d, ${}^{3}J$ = 4.7 Hz, 1H), 9.09–9.11 (m, 2H), 9.35 (d, ${}^{3}J$ = 4.7 Hz, 2H), 9.74 (s, 1H) ppm. 13 C NMR (100 MHz, CDCl₃): δ 14.3, 14.6, 14.8, 22.8, 23.7, 29.5, 29.9, 32.1, 34.8, 41.2, 53.6, 72.2, 72.3, 102.9, 111.8, 112.1, 121.0, 121.0, 121.4, 121.5, 122.9, 122.9, 126.6, 126.6, 127.3, 129.8, 130.0, 130.1, 130.2, 130.3, 130.4, 130.5, 130.8, 131.2, 131.3, 131.8, 132.2, 133.0, 135.1, 135.2, 161.1, 161.2 ppm. MS (MALDI): m/z 680.330 [M]*+. HRMS (ESI): calcd for C₄₆H₄₁N₄O₂ [M + H]⁺ 681.32240, found 681.32395.

BHP 11i (strap: 4Me, *t*-Bu-aryl; *meso*: H/*n*-Bu). Prepared from BHP **10c** (76 mg) and *n*-BuLi (0.12 mL, 2.5 M in *n*-hexane), column chromatography with DCM/*n*-hexane 40/60 \rightarrow 50/50. 62 mg, 76% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.37 (s, 2H), 0.02 (s, 6H), 0.14 (s, 6H), 1.10 (t, ³J = 7.4 Hz, 3H), 1.72 (m, 2H), 1.78 (s, 18H), 2.41–2.51 (m, 2H), 3.70 (d, ³J = 9.8 Hz, 2H), 3.75 (d, ³J = 9.8 Hz, 2H), 4.81–4.88 (m, 2H), 6.63 (d, ³J = 8.4 Hz, 2H), 7.62 (dd, ³J = 8.3 Hz, ⁴J = 2.5 Hz, 2H), 8.80 (d, ³J = 4.6 Hz, 2H), 8.83 (d, ³J = 4.7 Hz, 2H), 9.01–9.04 (m, 4H), 9.29 (d, ³J = 4.8 Hz, 2H), 9.68 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.5, 13.5, 14.3, 23.6, 31.8, 32.3, 34.7, 34.8, 40.9, 63.8, 102.2, 110.2, 113.7, 120.5, 125.8, 126.1, 128.0, 130.7, 130.8, 131.0, 131.0, 131.1, 131.4, 142.1, 156.6 ppm. MS (MALDI): *m/z* 821.500 [M]^{•+}. HRMS (ESI): calcd for C₅₆H₆₁N₄O₂ [M + H]⁺ 821.47890, found 821.47958.

BHP 11j (strap: 4Me, *t*-Oct-aryl; *meso*: H/Ph). Prepared from BHP **10c** (76 mg) and PhLi (0.12 mL, 1.9 M in dibutyl ether), column chromatography with DCM/*n*-hexane 40/60 → 70/30. 54 mg, 64% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.57 (s, 2H), 0.07 (s, 6H), 0.10 (s, 6H), 1.14 (s, 18H), 1.47 (s, 2H), 1.80 (d, ³*J* = 8.3 Hz, 12H), 3.69 (s, 4H), 6.59 (d, ³*J* = 8.4 Hz, 2H), 7.58 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 7.68–7.77 (m, 3H), 8.70 (d, ³*J* = 4.6 Hz, 2H), 8.75 (d, ³*J* = 4.7 Hz, 2H), 8.84 (d, ³*J* = 4.6 Hz, 2H), 9.01 (d, ⁴*J* = 2.5 Hz, 2H), 9.09 (d, ³*J* = 4.6 Hz, 2H), 9.80 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.6, 142.4, 141.1, 134.6, 131.5, 131.2, 131.2, 131.1, 131.1, 130.9, 130.9, 127.8, 127.3, 127.0, 126.7, 119.9, 114.6, 110.3, 103.2, 77.7, 77.6, 77.4, 77.0, 64.0, 57.7, 38.8, 33.2, 32.9, 32.7, 32.7, 32.6, 13.8, 13.7, 13.6 ppm. MS (MALDI): *m*/*z* 952.700 [M]^{*+}. HRMS (ESI): calcd for C₆₆H₇₃N₄O₂ [M + H]⁺ 953.57280, found 953.57257.

BHP 11k (strap: 4Me, *t*-Bu-aryl; *meso*: H/Ph). Prepared from BHP **10e** (88 mg) and PhLi (0.12 mL, 1.9 M in dibutyl ether), column chromatography with DCM/*n*-hexane 20/80 → 50/50. 57 mg, 65% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.57 (s, 2H), 0.03 (s, 6H), 0.12 (s, 6H), 1.75 (s, 18H), 3.69 (d, ³J = 1.1 Hz, 4H), 6.61 (d, ³J = 8.4 Hz, 2H), 7.60 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 7.72 (dd, ³J = 5.3 Hz, ⁴J = 2.1 Hz, 3H), 8.13 (s, 2H), 8.69 (d, ³J = 4.6 Hz, 2H), 8.76 (d, ³J = 4.7 Hz, 2H), 8.85 (d, ³J = 4.6 Hz, 2H), 9.02 (d, ⁴J = 2.5 Hz, 2H), 9.10 (d, ³J = 4.6 Hz, 2H), 9.80 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.7, 142.4, 142.3, 134.6, 131.6, 131.2, 131.2, 131.1, 131.1, 131.0, 130.9, 127.8, 127.0, 126.4, 125.8, 119.9, 114.5, 110.4, 103.2, 64.1, 35.0, 32.4, 13.8, 13.7 ppm. MS (MALDI): *m/z* 840.357 [M]^{•+}. HRMS (ESI): calcd for C₅₈H₅₇N₄O₂ [M + H]⁺ 841.44760, found 841.44959.

General Procedure E: *meso*-Bromination of BHPs. The BHP (100 μ mol, 1 equiv) was dissolved in chloroform (20 mL) and cooled to 5 °C. NBS was added in small portions over 2 min and the mixture stirred at 5 °C for 10 min. After filtration over a plug of silica the solvent was removed and the resulting solid recrystallized from chloroform/methanol to yield purple crystals. If a product mixture was obtained, the different brominated BHPs were resolved by column chromatography. BHPs 13a,b were reported before.¹²

BHP 13c (strap: 4Me, *t*-Bu-aryl; *meso*: 2 × Br). Prepared from BHP **10c** (76 mg) and NBS (37 mg, 0.21 mmol, 2 equiv). 91 mg, 97% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.04 (s, 2H), 0.23 (s, 12H), 1.75 (s, 18H), 3.80 (s, 4H), 6.63 (d, ³*J* = 8.5 Hz, 2H), 7.62 (dd, ³*J* = 8.5 Hz, ⁴*J* = 2.5 Hz, 2H), 8.71 (d, ³*J* = 4.8 Hz, 4H), 8.92 (d, ⁴*J* = 2.5 Hz, 2H), 9.31 (d, ³*J* = 4.8 Hz, 4H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.8, 32.2, 34.7, 53.6, 63.8, 110.2, 125.9, 136.6, 130.0, 130.9, 131.2, 132.1, 132.6, 142.2, 156.4 ppm. MS (MALDI): *m/z* 922.272 [M]^{•+}. HRMS (ESI): calcd for C₅₂H₅₁Br₂N₄O₂ [M + H]⁺ 921.23733, found 921.23844.

BHP 13d (strap: 2Me, *t*-Bu-aryl; *meso*: 2 × Br). Prepared from BHP 10d (73 mg) and NBS (37 mg, 0.21 mmol, 2 equiv). 89 mg, 99% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.96 (s, 2H), -0.47 (s, 6H), 1.76 (s, 18H), 2.46 (d, ³*J* = 13.4 Hz, 2H), 3.49 (d, ³*J* = 13.4 Hz, 2H), 3.66 (s, 2H), 7.01 (d, ³*J* = 8.4 Hz, 2H), 7.68 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.6 Hz, 2H), 8.64 (d, ³*J* = 4.8 Hz, 2H), 8.87 (d, ³*J* = 4.7 Hz, 2H), 8.97 (d, ⁴*J* = 2.5 Hz, 2H), 9.36 (d, ³*J* = 4.8 Hz, 2H), 9.38 (d, ³*J* = 4.7 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 158.9, 145.9, 134.1, 133.6, 132.1, 131.1, 130.8, 128.5, 127.1, 127.0, 121.8, 120.4, 115.0, 102.4, 72.4, 35.1, 32.3, 14.8 ppm. MS (MALDI): *m/z* 892.177 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₇Br₂N₄O₂ [M + H]⁺ 893.20603, found 893.20622.

BHP 13e (strap: 4Me, *t*-Oct-aryl; *meso*: 2 × Br). Prepared from BHP **10e** (88 mg) and NBS (37 mg, 0.21 mmol, 2 equiv). 99 mg, 97% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.12 (s, 2H), 0.22 (s, 12H), 1.14 (s, 18H), 1.52 (d, ³J = 1.7 Hz, 4H), 1.79 (s, 12H), 2.15 (s, 4H), 3.78 (s, 4H), 6.61 (d, ³J = 8.4 Hz, 2H), 7.59 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 8.68 (d, ³J = 4.8 Hz, 2H), 8.92 (d, ⁴J = 2.5 Hz, 2H), 9.31 (d, ³J = 4.8 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.5, 141.1, 132.7, 132.3, 131.4, 131.0, 129.9, 127.7, 126.9, 110.2, 77.7, 77.6, 77.4, 77.0, 63.9, 57.6, 38.8, 33.2, 32.8, 32.7, 32.7, 14.1, 1.4 ppm. MS (MALDI): *m/z* 1032.278 [M]^{•+}. HRMS (ESI): calcd for $C_{60}H_{67}Br_2N_4O_2$ [M + H]⁺ 1033.36253, found 1033.36225.

BHP 13f (strap: 2Me, *t*-Oct-aryl; *meso*: 2 × Br). Prepared from BHP 10f (85 mg) and NBS (37 mg, 0.21 mmol, 2 equiv). 96 mg, 96% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -3.53 (s, 2H), -0.57 (s, 6H), 1.17 (s, 18H), 1.84 (d, ³J = 13.4 Hz, 12H), 2.20 (q, ³J = 14.5 Hz, 4H), 2.34 (d, ³J = 13.5 Hz, 2H), 3.42 (d, ³J = 13.4 Hz, 2H), 3.52 (s, 2H), 6.96 (d, ³J = 8.4 Hz, 2H), 7.66 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 8.56 (d, ³J = 4.8 Hz, 2H), 8.79 (d, ³J = 4.8 Hz, 2H), 9.02 (d, ⁴J = 2.5 Hz, 2H), 9.30–9.33 (m, 4H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.5, 32.0, 32.5, 32.9, 33.0, 38.9, 57.5, 72.2, 102.0, 114.5, 120.1, 121.4, 126.6, 127.8, 129.1, 130.6, 130.7, 131.5, 133.3, 133.4, 133.9, 144.6, 158.6 ppm. MS (MALDI): *m*/*z* 1004.291 [M]*+. HRMS (ESI): calcd for C₅₈H₆₃Br₂N₄O₂ [M + H]⁺ 1005.33123, found 1005.33244.

BHP 13g (strap: 2Br, H-aryl; *meso*: 2 × Br). Prepared from BHP **10g** (75 mg) and NBS (37 mg, 0.21 mmol, 2 equiv). 88 mg, 97% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.87 (s, 2H), 2.36 (d, ³*J* = 14.4 Hz, 2H), 3.36 (d, ³*J* = 14.4 Hz, 2H), 4.04 (s, 2H), 7.12 (dd, ³*J* = 8.0 Hz, ⁴*J* = 1.2 Hz, 2H), 7.69 (td, ³*J* = 7.8 Hz, ⁴*J* = 1.8 Hz, 2H), 7.78 (td, ³*J* = 7.6 Hz, ⁴*J* = 1.3 Hz, 2H), 8.58 (d, ³*J* = 4.8 Hz, 2H), 8.91 (d, ³*J* = 4.8 Hz, 2H), 9.42 (d, ³*J* = 7.8 Hz, ⁴*J* = 1.8 Hz, 2H), 9.39 (d, ³*J* = 4.8 Hz, 2H), 9.42 (d, ³*J* = 4.8 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 160.7, 134.7, 134.6, 133.6, 132.9, 132.3, 132.1, 131.6, 130.7, 125.6, 123.9, 122.3, 115.0, 113.5, 103.6, 77.4, 73.6 ppm. MS (MALDI): *m*/*z* 907.838 [M]^{•+}. HRMS (ESI): calcd for C₄₀H₂₅Br₄N₄O₂ [M + H]⁺ 908.87055, found 908.87262.

BHP 13h (strap: 2Br, *t*-Oct-aryl; *meso*: 2 × Br). Prepared from BHP 10h (98 mg) and NBS (37 mg, 0.21 mmol, 2 equiv). 113 mg, 99% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -3.94 (s, 2H), 1.18 (s, 18H), 1.83 (s, 6H), 1.87 (s, 6H), 2.08 (d, ³*J* = 14.6 Hz, 2H), 2.14–2.29 (m, 4H), 3.27 (d, ³*J* = 14.6 Hz, 2H), 3.75 (s, 2H), 6.96 (d, ³*J* = 8.4 Hz, 2H), 7.66 (dd, ³*J* = 8.5 Hz, ⁴*J* = 2.5 Hz, 2H), 8.46 (d, ³*J* = 4.8 Hz, 2H), 8.75 (d, ³*J* = 4.8 Hz, 2H), 9.06 (d, ⁴*J* = 2.5 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 32.0, 32.5, 33.0, 33.1, 39.0, 57.6, 73.2, 102.8, 113.1, 114.4, 121.0, 124.9, 127.8, 129.5, 131.1, 131.4, 132.6, 133.2, 133.6, 134.1, 145.3, 158.1 ppm. MS (MALDI): *m/z* 1136.179 [M]^{•+}. HRMS (ESI): calcd for C₅₆H₅₇Br₄N₄O₂ [M + H]⁺ 1133.12095, found 1133.11982.

BHP 13i (strap: 4H, *t*-Oct-aryl; *meso*: 2 × Br). Prepared from BHP **10i** (82 mg) and NBS (37 mg, 0.21 mmol, 2 equiv). 97 mg, 99% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.04 (s, 2H), 1.14 (s, 18H), 1.80 (s, 12H), 2.16 (s, 4H), 3.41 (s, 4H), 3.45 (s, 4H), 6.69 (d, ³J = 8.4 Hz, 2H), 7.61 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 8.67 (d, ³J = 4.8 Hz, 4H), 8.99 (d, ⁴J = 2.5 Hz, 2H), 9.34 (d, ³J = 4.8 Hz, 4H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.9, 142.7, 134.1, 132.9, 132.4, 131.2, 127.9, 127.8, 123.8, 115.8, 114.7, 102.4, 77.2, 71.0, 57.5, 53.6, 38.7, 33.0, 32.4 ppm. MS (MALDI): *m/z* 976.226 [M]^{•+}. HRMS

(ESI): calcd for $C_{56}H_{59}Br_2N_4O_2\ [M + H]^+$ 977.29993, found 977.30318.

BHP 14a (strap: 2Me, H-aryl; meso: n-Bu/Br). Prepared from BHP 11h (68 mg) and NBS (19 mg, 105 µmol, 1 equiv). 71 mg, 96% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.58 (s, 2H), -0.52 (s, 3H), -0.37 (s, 3H), 1.11 (t, ${}^{3}J$ = 7.4 Hz, 3H), 1.75 (q, ${}^{3}J$ = 7.4 Hz, 2H), 2.38–2.51 (m, 2H), 2.63 (d, ${}^{3}J$ = 12.4 Hz, 2H), 3.56 (app t, ${}^{app}J$ = 11.3 Hz, 2H), 3.73 (s, 1H), 3.79 (s, 1H), 4.67-4.75 (m, 2H), 7.09 (app t, ^{app}J = 1.4 Hz, 1H), 7.11 (app t, ^{app}J = 1.4 Hz, 1H), 7.68 (app tt, ${}^{3}J = 1.6 \text{ Hz}, {}^{\text{app}}J = 7.7 \text{ Hz}, 2\text{H}), 7.74 \text{ (app tt, } {}^{3}J = 1.6 \text{ Hz}, {}^{\text{app}}J = 7.5 \text{ Hz},$ 2H), 8.65-8.68 (m, 2H), 8.90-8.93 (m, 3H), 8.95 (d, ${}^{3}I = 4.7$ Hz, 1H), 9.24 (dd, ${}^{3}J$ = 4.7 Hz, ${}^{4}J$ = 1.4 Hz, 2H), 9.38 (d, ${}^{3}J$ = 4.8 Hz, 1H), 9.40 (d, ${}^{3}J$ = 4.8 Hz, 1H) ppm. ${}^{13}C$ NMR (100 MHz, CDCl₃): δ 14.3, 14.7, 14.9, 23.7, 29.9, 34.5, 41.0, 72.1, 72.2, 100.5, 113.2, 113.4, 120.7, 120.8, 121.3, 121.9, 122.0, 122.8, 122.8, 126.9, 127.1, 127.6, 130.2, 130.2, 130.2, 130.7, 130.7, 130.8, 131.1, 131.1, 131.2, 131.7, 132.3, 133.0, 133.3, 134.7, 134.8, 143.4, 143.7, 144.0, 147.4, 161.0, 161.0 ppm. MS (MALDI): *m*/*z* 758.249 [M]^{•+}. HRMS (ESI): calcd for $C_{46}H_{40}BrN_4O_2 [M + H]^+$ 759.23292, found 759.23368.

BHP 14b (strap: 4Me, H-aryl; *meso: n*-Bu/Br). Prepared from BHP **11e** (71 mg) and NBS (19 mg, 105 μ mol, 1 equiv). 73 mg, 94% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –1.94 (s, 2H), 0.11 (s, 6H), 0.32 (s, 6H), 1.08 (t, ³J = 7.4 Hz, 2H), 1.62–1.79 (m, 2H), 2.30–2.48 (m, 2H), 3.80 (d, ³J = 9.9 Hz, 2H), 3.84 (d, ³J = 9.8 Hz, 2H), 4.73 (d, ³J = 7.2 Hz, 2H), 6.71 (dd, ³J = 7.9 Hz, ⁴J = 1.2 Hz, 2H), 7.51–7.64 (m, 4H), 8.74 (d, ⁴J = 2.4 Hz, 2H), 8.76 (d, ⁴J = 2.4 Hz, 1H), 8.88 (dd, ³J = 7.2 Hz, ⁴J = 1.2 Hz, 2H), 9.19 (d, ³J = 4.7 Hz, 2H), 9.32 (d, ³J = 4.8 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 158.7, 145.9, 145.5, 132.3, 131.6, 131.6, 131.4, 131.2, 131.0, 130.7, 130.1, 128.5, 128.4, 121.0, 119.3, 114.3, 110.7, 99.6, 63.6, 40.6, 34.4, 23.6, 14.3, 14.0, 13.6 ppm. MS (MALDI): *m*/*z* 786.417 [M]^{•+}. HRMS (ESI): calcd for C₄₈H₄₄BrN₄O₂ [M + H]⁺ 787.26422, found 787.26544.

BHP 14c (strap: 4Me, *t*-Bu-aryl; *meso*: (4-*t*-Bu)Ph/Br). Prepared from BHP 11b (90 mg) and NBS (19 mg, 105 μmol, 1 equiv). 97 mg, 99% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.05 (s, 2H), 0.08 (s, 6H), 0.36 (s, 6H), 1.59 (s, 18H), 1.75 (s, 9H), 3.74 (d, ³J = 9.9 Hz, 2H), 3.83 (d, ³J = 9.9 Hz, 2H), 6.64 (d, ³J = 8.4 Hz, 2H), 7.61 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 7.71-7.79 (m, 2H), 8.04 (d, ³J = 7.7 Hz, 2H), 8.68 (d, ³J = 4.7 Hz, 2H), 8.70 (d, ³J = 4.7 Hz, 2H), 8.79 (d, ³J = 4.7 Hz, 2H), 8.97 (d, ⁴J = 2.5 Hz, 2H), 9.38 (d, ³J = 4.8 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.7, 150.8, 146.9, 146.6, 146.3, 145.5, 142.3, 138.6, 134.4, 132.5, 131.9, 131.6, 131.5, 131.2, 131.0, 130.8, 126.5, 126.1, 124.2, 120.6, 115.5, 110.4, 100.5, 64.1, 35.2, 34.9, 32.4, 32.0, 14.3, 13.7 ppm. MS (MALDI): *m/z* 976.369 [M]^{•+}. HRMS (ESI): calcd for C₆₂H₆₄BrN₄O₂ [M + H]⁺ 975.42072, found 975.42239.

BHP 14d (strap: 4Me, *t*-Bu-aryl; *meso*: H/Br). Prepared from BHP **10c** (75 mg) and NBS (22 mg, 125 μmol, 1.2 equiv). column chromatography with chloroform/*n*-hexane 40/60 \rightarrow 70/30. 52 mg, 62% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.49 (s, 2H), 0.04 (s, 6H), 0.23 (s, 6H), 1.77 (s, 18H), 3.67 (d, ³*J* = 10.0 Hz, 2H), 3.74 (d, ³*J* = 10.0 Hz, 2H), 6.61 (d, ³*J* = 8.4 Hz, 2H), 7.62 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 8.78 (d, ³*J* = 3.2 Hz, 2H), 8.80 (d, ³*J* = 3.4 Hz, 2H), 8.99 (d, ⁴*J* = 2.5 Hz, 2H), 9.04 (d, ³*J* = 4.6 Hz, 2H), 9.44 (d, ³*J* = 4.6 Hz, 2H), 9.77 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.4, 13.7, 32.2, 34.8, 63.8, 101.7 103.9, 110.2, 114.9, 125.7, 126.4, 130.4, 130.7, 131.0, 131.1, 131.4, 131.7, 131.9, 142.2, 156.4 ppm. MS (MALDI): *m*/*z* 844.321 [M]^{•+}. HRMS (ESI): calcd for C₅₂H₅₂BrN₄O₂ [M + H]⁺ 843.32697, found 843.32682. BHPs **10c** (11%) and **13c** (25%) were isolated as side products.

BHP 14e (strap: 4Me, H-aryl; *meso*: H/Br). Prepared from BHP **10a** (64 mg) and NBS (22 mg, 125 μ mol, 1.2 equiv). column chromatography with chloroform/*n*-hexane 40/60 \rightarrow 70/30. 45 mg, 59% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.52 (s, 2H), -0.02 (s, 6H), 0.23 (s, 6H), 3.70 (d, ³J = 9.7 Hz, 2H), 3.76 (d, ³J = 9.7 Hz, 2H), 6.69 (d, ³J = 7.9 Hz, 2H),), 7.50-7.65 (m, 4H) 8.73-8.86 (m, 4H), 8.87-8.97 (m, 2H), 9.06 (d, ³J = 4.8 Hz, 2H), 9.39-9.50 (m, 2H), 9.79 (s, 1H) ppm. ¹3C NMR (100 MHz, CDCl₃): δ 158.6, 132.0, 131.6, 131.1, 130.8, 130.5, 130.2, 128.2, 119.4, 114.4, 110.8, 104.0, 63.6, 13.8, 13.5 ppm. MS (MALDI): *m/z* 730.225 [M]^{•+}.

HRMS (ESI): calcd for $C_{44}H_{36}BrN_4O_2$ [M + H]⁺ 731.20162, found 731.20411. BHPs **10a** (12%) and **13a** (26%) were isolated as side products.

General Procedure F: β -Borylation of BHPs. The BHP (0.2 mmol), dtbpy (11 mg, 0.03 mmol), [Ir(cod)OMe]₂ (11 mg, 0.02 mmol), and (Bpin)₂ were dissolved in absolute THF under an inert atmosphere and heated to reflux. After filtration over a plug of silica, the solvent was evaporated and the crude product was purified by preparative recycling GPC to yield the borylated BHPs. The product ratio can be influenced by variation of the amount of (Bpin)₂ and the reaction time. BHPs **15a**-e were reported before.¹²

BHP 15f (strap: 4Me, t-Bu-aryl; meso: (4-t-Bu)Ph/H; β : Bpin). Prepared from BHP 11b (180 mg) and (Bpin)₂ (100 mg, 0.4 mol, 2 equiv) by heating for 24 h. 119 mg, 55% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –2.30 (s, 2H), 0.06 (d, ³J = 16.7 Hz, 6H), 0.24 (d, ³*J* = 19.8 Hz, 6H), 1.21 (s, 12H), 1.22 (s, 18), 1.66 (s, 6H), 1.70 (s, 6H), 1.75 (d, ${}^{3}J$ = 2.7 Hz, 18H), 3.64 (d, ${}^{3}J$ = 9.9 Hz, 1H), 3.70 (d, ${}^{3}J$ = 9.9 Hz, 1H), 3.77 (app t, $^{app}J = 9.7$ Hz, 2H), 6.58 (d, $^{3}J = 8.4$ Hz, 1H), 6.62 (d, ${}^{3}J = 8.4$ Hz, 1H), 7.56–7.63 (m, 2H), 7.76 (app t, ${}^{app}J = 1.8$ Hz, 1H), 8.68 (q, ${}^{3}J$ = 4.7 Hz, 2H), 8.72 (q, ${}^{3}J$ = 4.7 Hz, 2H), 8.83 (d, ${}^{3}J = 4.6$ Hz, 1H), 9.00 (d, ${}^{4}J = 2.5$ Hz, 1H), 9.02 (d, ${}^{4}J = 2.5$ Hz, 1H), 9.15 (d, ³J = 4.6 Hz, 1H), 9.42 (s, 1H), 10.39 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₂): δ 13.3, 13.3, 13.9, 14.0, 24.7, 24.8, 24.9, 24.9, 24.9, 25.0, 25.4, 25.5, 26.1, 26.4, 26.6, 26.8, 27.0, 27.2, 27.3, 27.7, 29.0, 29.8, 29.9, 31.6, 31.8, 32.2, 32.3, 34.8, 35.1, 63.7, 63.8, 77.4, 82.9, 82.9, 83.1, 84.2, 104.3, 110.0, 110.1, 113.4, 114.8, 120.9, 125.7, 125.9, 126.1, 126.2, 129.5, 130.8, 130.8, 130.8, 130.9, 130.9, 131.1, 131.1, 131.2, 131.3, 141.1, 142.0, 142.1, 146.2, 148.8, 156.6 ppm. MS (MALDI): m/ z 1078.623 [M]^{•+}. HRMS (ESI): calcd for $C_{72}H_{84}BN_4O_4$ [M + H]⁺ 1079.65801, found 1079.65867. BHPs 11b (26%) and 15g (14%) were isolated as side products.

BHP 15g (strap: 4Me, *t*-Bu-aryl; *meso*: (4-*t*-Bu)Ph/H; β: 2 × Bpin). Prepared from BHP 11b (180 mg) and (Bpin)₂ (400 mg, 1.6 mol, 8 equiv) by heating for 4 d. 184 mg, 77% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –1.99 (s, 2H), –0.15 (s, 6H), 0.09 (s, 2H), 0.59 (s, 6H), 1.21 (s, 9H), 1.68 (s, 12H), 1.71 (s, 12H), 1.76 (s, 18H), 3.66 (d, ³J = 9.9 Hz, 2H), 3.91 (d, ³J = 9.9 Hz, 2H), 6.61 (d, ³J = 8.5 Hz, 2H), 7.59 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 7.75 (app t, ^{app}J = 1.8 Hz, 1H), 8.67 (s, 4H), 9.01 (d, ⁴J = 2.5 Hz, 2H), 9.42 (s, 2H), 10.85 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 1.2, 13.1, 14.6, 24.7, 25.1, 25.5, 25.5, 29.9, 31.6, 31.8, 32.3, 34.7, 35.1, 63.7, 77.4, 82.9, 84.1, 105.7, 109.9, 113.9, 120.6, 120.9, 126.0, 126.2, 129.4, 130.6, 130.9, 130.9, 131.3, 131.6, 140.9, 141.9, 142.5, 148.9, 156.8 ppm. MS (MALDI): m/z 1204.738 [M]^{•+}. HRMS (ESI): calcd for C₇₈H₉₅B₂N₄O₆ [M + H]⁺ 1205.74322, found 1205.74606. BHPs **11b** (4%) and 15f (12%) were isolated as side products.

2-Bromo-5,10,15,20-tetrakis(4-tert-butylphenyl)porphyrin (16b). 5,10,15,20-Tetrakis(4-tert-butylphenyl)porphyrin (1.00 g, 1.19 mmol) was dissolved in 1,2-dichlorobenzene (200 mL) and heated to 160 °C. NBS (212 mg, 1.19 mmol) was added and the solution stirred at 160 °C for 45 min. A 150 mL amount of the solvent was removed, and MeOH (300 mL) was added. The precipitate was isolated and purified by column chromatography (silica, *n*-hexane/dichloromethane 1/1). The obtained solid was recrystallized from chloroform/methanol to yield purple crystals of 16b. 764 mg, 70% yield; mp >300 °C. ¹H NMR (400 MHz, CDCl₂): δ –2.82 (s, 2H), 1.59 (s, 9H), 1.60 (s, 9H), 1.61 (s, 9H), 1.62 (s, 9H), 7.71-7.80 (m, 8H), 7.97-8.03 (m, 2H),), 8.08-8.17 (m, 6H), 8.79 (s, 1H), 8.80-8.96 (m, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 150.6, 150.5, 150.3, 140.7, 139.6, 139.5, 139.2, 136.6, 135.0, 134.8, 134.7, 123.9, 123.8, 123.7, 123.7, 121.8, 120.4, 120.1, 119.9, 83.8, 35.1, 35.0, 35.0, 31.9, 31.9, 25.4 ppm. MS (MALDI): m/z 916.347 [M]^{•+}. HRMS (ESI): calcd for $C_{60}H_{62}BrN_4$ $[M + H]^+$ 917.41524, found 917.41752.

2-(4,4,5,5-Tetramethyl[1,3,2]dioxaborolan-2-yl)-5,10,15,20tetrakis(4-tolyl)porphyrin (17). Porphyrin 16b (350 mg, 0.38 mmol), (Bpin)₂ (260 mg, 0.95 mmol) and KOAc (370 mg, 3.79 mmol) were dissolved in toluene (100 mL) and water (20 mL) and degassed with an argon stream in an ultrasonic bath for 15 min. Pd(dppf)Cl₂ (31 mg, 0.038 mmol) was added and the mixture was degassed for a further 20 min. Then the reaction mixture was heated to 110 °C for 6 h. After it was cooled to room temperature, the organic phase was filtered over a plug of silica and fully eluted with ethyl acetate. After evaporation of all solvents, the residue was purified by column chromatography (silica, *n*-hexane/dichloromethane 2/1 \rightarrow 0/1). The product was recrystallized from chloroform/methanol to yield a purple solid. 304 mg, 83% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.57 (s, 2H), 1.59 (s, 9H), 1.62 (s, 9H), 1.62 (s, 9H), 1.64 (s, 9H), 7.75–7.83 (m, 8H), 8.12–8.27 (m, 8H), 8.64 (d, ³*J* = 4.9 Hz, 1H), 8.77 (d, ³*J* = 4.8 Hz, 1H), 8.85 (d, ³*J* = 5.3 Hz, 4H), 9.14 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 150.6, 150.5, 150.3, 140.7, 139.6, 139.5, 139.2, 136.6, 135.0, 134.8, 134.7, 123.9, 123.8, 123.7, 123.7, 121.8, 120.4, 120.1, 119.9, 83.8, 35.1, 35.0, 35.0, 31.9, 31.9, 25.4 ppm. MS (MALDI): *m*/*z* 964.566 [M]⁺. HRMS (ESI): calcd for C₆₆H₇₄BN₄O₂ [M + H]⁺ 965.58993, found 965.59112.

 $\beta_{,\beta}$ -Dimer 18. Porphyrin 16a³² (94 mg, 0.125 mmol), porphyrin 15a (89 mg, 0.100 mmol), and Ba(OH)₂ (315 mg, 1.0 mmol) were dissolved in toluene (100 mL) and water (20 mL) and degassed with an argon stream in an ultrasonic bath for 15 min. $Pd(PPh_3)_4$ (10 mg, 0.01 mmol) was added and the mixture degassed for further 20 min. Then the reaction mixture was heated to 110 °C for 6 h. After it was cooled to room temperature, the organic phase was filtered over a plug of silica and fully eluted with ethyl acetate. After evaporation of all solvents, the residue was purified by column chromatography (silica, nhexane/dichloromethane $2/1 \rightarrow 0/1$). 125 mg, 83% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.75 (s, 2H), -2.42 (s, 3H), -0.08 (s, 6H), 0.11 (s, 3H), 0.15 (s, 3H), 1.74 (s, 9H), 1.77 (s, 9H), 2.47 (s, 3H), 2.69 (s, 6H), 2.74 (s, 3H), 3.60 (s, 2H), 3.63 (d, ³*J* = 10.1 Hz, 2H), 3.69 (d, ${}^{3}J$ = 10.1 Hz, 2H), 4.78 (d, ${}^{3}J$ = 7.7 Hz, 2H), 6.51 (d, ${}^{3}J = 8.5, 2H), 6.60 (d, {}^{3}J = 8.5 Hz, 2H), 7.04 (d, {}^{3}J = 7.5 Hz, 2H),$ 7.38–7.58 (m, 18H), 7.58–7.66 (m, 4H), 8.11 (d, ${}^{3}J$ = 7.7 Hz, 2H), 8.18 (d, ³*J* = 7.8 Hz, 4H), 8.37 (s, 1H), 8.50 (d, ³*J* = 5.3 Hz, 2H), 8.77 $(d, {}^{3}I = 4.8 \text{ Hz}, 1\text{H}), 8.85 (d, {}^{3}I = 4.5 \text{ Hz}, 2\text{H}), 8.88 (d, {}^{3}I = 4.8 \text{ Hz}, 2\text{H})$ 1H), 8.91 (d, ${}^{3}J$ = 4.6 Hz, 2H), 8.97 (d, ${}^{3}J$ = 4.8 Hz, 2H), 8.99–9.03 (m, 2H), 9.10 (d, ${}^{3}J$ = 2.5 Hz, 1H), 9.16 (d, ${}^{3}J$ = 4.6 Hz, 1H), 9.20– 9.23 (m, 2H), 9.64 (s, 1H), 9.95 (s, 1H). ${}^{13}C$ NMR (100 MHz, CDCl₃): δ 156.4, 156.3, 142.1, 141.9, 139.9, 139.5, 139.3, 138.1, 137.5, 137.5, 137.3, 136.9, 135.3, 134.8, 134.8, 134.7, 133.9, 131.1, 131.0, 130.9, 130.9, 130.8, 130.6, 130.5, 127.7, 127.5, 127.3, 127.2, 126.8, 126.2, 126.1, 126.0, 125.6, 125.0, 122.0, 120.5, 120.2, 120.1, 113.9, 112.7, 110.0, 109.8, 103.5, 102.7, 68.1, 63.6, 63.4, 34.8, 34.8, 32.3, 32.2, 32.2, 31.9, 29.9, 21.7, 21.7, 21.5, 20.2, 14.3, 13.6, 13.4, 13.3, 13.3 ppm. MS (MALDI): m/z 1332.815 [M]⁺. HRMS (ESI): calcd for $C_{100}H_{89}N_8O_2$ [M + H]⁺ 1433.71030, found 1433.71399.

β,meso-Dimer 19. Porphyrin 16b (96 mg, 0.100 mmol), porphyrin 14e (105 mg, 0.125 mmol), and Ba(OH)₂ (315 mg, 1.0 mmol) were dissolved in toluene (100 mL) and water (20 mL) and degassed with an argon stream in an ultrasonic bath for 15 min. $Pd(PPh_3)_4$ (10 mg, 0.01 mmol) was added and the mixture degassed for further 20 min. Then the reaction mixture was heated to 110 °C for 6 h. After it was cooled to room temperature, the organic phase was filtered over a plug of silica and fully eluted with ethyl acetate. After evaporation of all solvents, the residue was purified by column chromatography (silica, *n*-hexane/dichloromethane $2/1 \rightarrow 0/1$). 125 mg, 78% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -2.66 (s, 2H), -2.30 (s, 2H), -1.14 (s, 9H), -0.10 (s, 6H), 0.24 (s, 6H), 1.37 (s, 9H), 1.58 (s, 9H), 1.67 (s, 9H), 3.63 (d, ${}^{3}J$ = 10.1 Hz, 2H), 3.70 (d, ${}^{3}J$ = 10.1 Hz, 2H), 5.19 (d, ${}^{3}J$ = 8.2 Hz, 1H), 6.63 (dd, ${}^{3}J$ = 8.3 Hz, ${}^{4}J$ = 2.2 Hz, 2H), 6.96 (d, ³J = 8.2 Hz, 2H), 7.47-7.57 (m, 4H), 7.68 (d, ³J = 8.4 Hz, 2H), 7.73 (d, ${}^{3}J$ = 8.4 Hz, 2H), 7.84 (d, ${}^{3}J$ = 7.8 Hz 2H), 8.15 (d, ${}^{3}J$ = 8.4 Hz, 2H), 8.26 (d, ${}^{3}J$ = 8.2 Hz, 2H), 8.37 (d, ${}^{3}J$ = 4.9 Hz, 1H), 8.39 (d, ${}^{3}J$ = 8.4 Hz, 2H), 8.47 (d, ${}^{3}J$ = 4.7 Hz, 2H), 8.48 (d, ${}^{3}J = 4.7$ Hz, 2H), 8.69 (d, ${}^{3}J = 4.8$ Hz, 1H), 8.84 (d, ${}^{3}J = 4.6$ Hz, 2H), 8.90-8.97 (m, 4H), 9.02 (d, ${}^{3}J$ = 4.8 Hz, 1H), 9.06 (d, ${}^{3}J$ = 4.8 Hz, 1H), 9.09 (d, ${}^{3}J$ = 4.6 Hz, 2H), 9.60 (s, 1H), 9.80 (s, 1H) ppm. ${}^{13}C$ NMR (100 MHz, CDCl₃): δ 158.7, 150.7, 150.6, 150.4, 147.4, 139.8, 139.4, 139.3, 136.4, 134.8, 134.8, 134.6, 133.0, 132.0, 131.0, 131.0, 130.5, 129.8, 129.2, 128.4, 127.8, 123.9, 123.7, 123.6, 122.5, 120.6, 120.5, 120.3, 119.4, 113.9, 110.7, 102.9, 63.6, 53.6, 35.1, 35.0, 34.9, 32.1, 31.9, 31.9, 31.8, 31.7, 29.9, 28.8, 13.7, 13.3. MS (MALDI): m/z

1204.738 [M]^{•+}. MS (MALDI): m/z 1488.759 [M]^{•+}. HRMS (ESI): calcd for $C_{104}H_{97}N_8O_2$ [M + H]⁺ 1489.77290, found 1489.77454.

General Procedure G: Metalation of BHPs. The BHP (0.1 mmol, 1 equiv) was dissolved in an appropriate solvent (30 mL), the metal salt (0.3 mmol, 3 equiv) was added, and the mixture was heated to reflux until no remaining starting material was detected by TLC or UV/vis. The solvent was removed and the remaining solid redissolved in chloroform. After filtration through a plug of silica and evaporation of the solvent, the porphyrin was recrystallized from chloroform/ methanol. All metalation reactions proceeded with yields >97%.

All metallo-BHPs M-X were prepared from the corresponding freebase BHP X: for Ni-BHPs, $(CH_2Cl)_2$ (100 mL), $Ni(acac)_2$ (78 mg); for Zn-BHPs, $CHCl_3$ (100 mL), $Zn(OAc)_2$ (66 mg, dissolved in 2 mL of MeOH); for Pd-BHPs, $(CH_2Cl)_2$ (100 mL), $Pd(OAc)_2$ (67 mg); for Mg-BHPs, $CHCl_3$ (100 mL), NEt_3 (0.5 mL), $MgBr_2 \cdot OEt_2$ (78 mg); for Cu-BHPs, $CHCl_3$ (100 mL), $Cu(OAc)_2$ (60 mg, dissolved in 1 mL of MeOH); for In-BHPs, glacial acetic acid (100 mL), NaOAc(136 mg, 1 mmol), $InCl_3$ (66 mg); for V-BHPs, quinoline (100 mL), $VO(acac)_2$ (79 mg).

The metal complexes of the BHPs 10a (Ni, Pd, Zn, Mg, Cu) and 10b (Ni) were reported before.¹²

BHP Zn-10b (strap: 2Me, H-aryl; *meso*: 2 × H) prepared from **10b** (66 mg). 68 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.78 (s, 6H), 2.05 (d, ³J = 13.6 Hz, 2H), 3.24 (s, 2H), 3.29 (d, ³J = 13.6 Hz, 3H), 7.09 (dd, ³J = 8.0 Hz, ⁴J = 1.2 Hz, 2H), 7.68 (dd, ³J = 7.8 Hz, ⁴J = 1.7 Hz, 2H), 7.80 (dd, ³J = 7.5 Hz, ⁴J = 1.2 Hz, 2H), 9.20 (d, ³J = 4.4 Hz, 2H), 9.06 (dd, ³J = 7.4 Hz, ⁴J = 1.8 Hz, 2H), 9.20 (d, ³J = 4.5 Hz, 2H), 9.30 (d, ³J = 4.4 Hz, 2H), 9.32 (d, ³J = 4.5 Hz, 2H), 10.06 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 1.2, 14.3, 14.4, 22.9, 29.5, 29.9, 32.1, 71.9, 77.4, 105.8, 113.0, 120.6, 121.7, 123.3, 126.3, 130.0, 130.3, 130.6, 131.6, 132.5, 132.7, 132.9, 135.9, 148.9, 149.0, 149.5, 151.0, 161.2 ppm. MS (MALDI): *m/z* 686.158 [M]^{•+}. HRMS (ESI): calcd for C₄₂H₃₀N₄O₂Zn [M + H]^{•+} 686.16547, found 686.16668.

BHP In-10b (strap: 2Me, H-aryl; *meso*: 2 × H) prepared from 10b (66 mg). 76 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.80 (s, 6H), 2.11 (d, ³J = 12.7 Hz, 2H), 3.14 (s, 2H), 3.22 (d, ³J = 12.7 Hz, 2H), 7.04 (dd, ³J = 7.9 Hz, ⁴J = 1.1 Hz, 2H), 7.71 (app dt, ⁴J = 1.8 Hz, ^{app}J = 7.8 Hz, 2H), 7.81 (app dt, ⁴J = 1.3 Hz, ^{app}J = 7.7 Hz, 2H), 8.91 (d, ³J = 4.5 Hz, 2H), 9.12 (dd, ³J = 7.4 Hz, ⁴J = 1.8 Hz, 2H), 9.16 (d, ³J = 4.5 Hz, 2H), 9.45 (d, ³J = 4.6 Hz, 2H), 9.49 (d, ³J = 4.6 Hz, 2H), 10.31 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.4, 71.3, 105.8, 115.3, 120.6, 121.2, 123.1, 126.5, 130.2, 130.5, 130.9, 132.3, 132.9, 133.1, 133.7, 134.9, 148.4, 148.5, 150.0, 151.1, 160.4 ppm. MS (MALDI): *m/z* 772.100 [M]^{•+}. HRMS (ESI): calcd for C₄₂H₃₀InN₄O₂ [M - Cl]⁺ 737.14021, found 737.14033.

BHP Pd-10c (strap: 4Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10c** (77 mg). 86 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.02 (s, 12H), 1.76 (s, 18H), 3.65 (s, 4H), 6.62 (d, ³*J* = 8.4 Hz, 2H), 7.61 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 8.92 (d, ³*J* = 4.7 Hz, 4H), 9.00 (d, ⁴*J* = 2.5 Hz, 2H), 9.09 (d, ³*J* = 4.7 Hz, 4H), 9.05 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.4, 32.2, 34.8, 64.1, 77.4, 105.8, 110.6, 125.3, 126.3, 130.6, 130.7, 131.0, 131.0, 131.3, 140.0, 142.2, 142.4, 156.5 ppm. MS (MALDI): *m/z* 866.227 [M]^{*+}. HRMS (ESI): calcd for C₅₂H₅₀N₄NaO₂Pd [M + Na]⁺ 891.28608, found 891.28868.

BHP In-10c (strap: 4Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10c** (77 mg). 90 mg, 97% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.26 (s, 12H), 1.79 (s, 18H), 3.43 (s, 4H), 6.53 (d, ⁴*J* = 8.2 Hz, 2H), 7.63 (dd, ³*J* = 8.2 Hz, ⁴*J* = 2.5 Hz, 2H), 8.94 (d, ³*J* = 4.5 Hz, 4H), 9.13 (d, ⁴*J* = 2.5 Hz, 2H), 9.89 (d, ³*J* = 4.5 Hz, 4H), 10.24 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.1, 14.3, 22.8, 29.9, 31.7, 32.2, 34.8, 63.1, 105.1, 109.7, 117.5, 125.6, 126.5, 130.2, 130.5, 131.1, 132.5, 132.6, 142.3, 148.3, 152.0, 156.0 ppm. MS (MALDI): *m/z* 912.311 [M]^{•+}. HRMS (ESI): calcd for C₅₂H₅₀InN₄O₂ [M - Cl]⁺ 877.29671, found 877.29659.

BHP Zn-10c (strap: 4Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10c** (77 mg). 81 mg, 97% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.12 (s, 12H), 1.77 (s, 18H), 3.52 (s, 4H), 6.57 (d, ³J = 8.4 Hz, 2H), 7.60 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 8.99 (d, ³J = 4.5

Hz, 4H), 9.08 (d, ³*J* = 2.5 Hz, 2H), 9.25 (d, ³*J* = 4.5 Hz, 4H), 10.04 (s, 2H) ppm. Due to limited solubility in common deuterated solvents, ¹³C NMR data are not available. MS (MALDI): m/z 826.379 [M]^{•+}. HRMS (ESI): calcd for C₅₂H₅₁N₄O₂Zn [M + H]⁺ 827.32980, found 827.32918.

BHP Ni-10c (strap: 4Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10c** (77 mg). 79 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.40 (s, 12H), 1.69 (s, 18H), 4.06 (s, 4H), 6.71 (d, ³*J* = 8.4 Hz, 2H), 7.59 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 8.79 (d, ³*J* = 4.7 Hz, 4H), 8.87 (d, ⁴*J* = 2.5 Hz, 2H), 8.94 (d, ³*J* = 4.7 Hz, 4H), 9.51 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.7, 142.3, 142.2, 140.8, 132.6, 132.4, 132.1, 131.6, 128.6, 127.6, 126.4, 112.1, 110.3, 103.8, 64.2, 51.0, 34.8, 32.3, 32.3, 14.6 ppm. MS (MALDI): *m/z* 820.307 [M]^{•+}. HRMS (ESI): calcd for C₅₂H₅₀N₄NaNiO₂ [M + Na]⁺ 843.31795, found 843.31916.

BHP Ni-10d (strap: 2Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10d** (73 mg). 79 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.43 (s, 6H), 1.70 (s, 18H), 2.86 (d, ³*J* = 13.1 Hz, 2H), 3.76 (d, ³*J* = 13.1 Hz, 2H), 4.23 (s, 2H), 7.06 (d, ³*J* = 8.4 Hz, 2H), 7.66 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.6 Hz, 2H), 8.71 (d, ³*J* = 4.7 Hz, 2H), 8.88 (d, ⁴*J* = 2.6 Hz, 2H), 8.99 (d, ³*J* = 4.7 Hz, 2H), 9.02 (d, ³*J* = 4.7 Hz, 2H), 9.05 (d, ³*J* = 4.7 Hz, 2H), 9.59 (s, 2H) ppm. ¹³C NMR (150 MHz, CDCl₃): δ 14.9, 32.1, 34.9, 72.9, 104.1, 111.7, 119.8, 122.8, 126.6, 127.8, 129.7, 131.1, 131.3, 132.3, 132.9, 141.4, 141.6, 142.9, 145.6, 158.6 ppm. MS (MALDI): *m/z* 792.216 [M]^{*+}. HRMS (ESI): calcd for C₅₀H₄₆N₄Ni₁O₂ [M]⁺ 792.29742, found 792.29688.

BHP Pd-10d (strap: 2Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from 10d (73 mg). 83 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.66 (s, 6H), 1.75 (s, 18H), 2.28 (d, ³J = 13.6 Hz, 2H), 3.46 (d, ³J = 13.6 Hz, 2H), 3.58 (s, 2H), 7.04 (d, ³J = 8.2 Hz, 2H), 7.68 (dd, ³J = 8.2 Hz, ⁴J = 2.0 Hz, 2H), 8.79 (d, ³J = 4.8 Hz, 2H), 9.02 (d, ⁴J = 2.0 Hz, 2H), 9.09 (d, ³J = 4.5 Hz, 2H), 9.12 (d, ³J = 4.5 Hz, 2H), 9.17 (d, ³J = 4.8 Hz, 2H), 9.97 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.5, 29.9, 32.1, 35.0, 72.5, 106.2, 114.1, 120.9, 121.4, 126.7, 128.2, 130.5, 130.6, 131.5, 131.9, 132.1, 133.8, 140.2, 140.4, 140.6, 142.1, 146.1, 185.9 ppm. MS (MALDI): *m*/*z* 838.205 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₆N₄O₂Pd [M]⁺ 840.26556, found 840.26501.

BHP Zn-10d (strap: 2Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10d** (73 mg). 79 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.80 (s, 6H), 1.78 (s, 18H), 1.99 (d, ³*J* = 13.4 Hz, 2H), 3.22 (s, 2H), 3.27 (d, ³*J* = 13.4 Hz, 2H), 7.01 (d, ³*J* = 8.4 Hz, 2H), 7.68 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.6 Hz, 2H), 8.91 (d, ³*J* = 4.5 Hz, 2H), 9.12 (d, ⁴*J* = 2.6 Hz, 2H), 9.19 (d, ³*J* = 4.5 Hz, 2H), 9.29 (d, ³*J* = 4.5 Hz, 2H), 9.33 (d, ³*J* = 4.5 Hz, 2H), 10.06 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 1.2, 14.3, 29.9, 32.2, 35.0, 72.0, 105.7, 113.6, 120.5, 120.9, 126.2, 126.4, 127.9, 130.4, 131.7, 132.3, 132.9, 132.9, 135.1, 146.1, 148.9, 149.0, 149.5, 151.1, 159.0 ppm. MS (MALDI, positiv): *m/z* 798.315 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₆N₄O₂Zn [M]⁺ 798.29122, found 798.29111.

BHP Mg-10d (strap: 2Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10d** (73 mg). 76 mg, 97% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.82 (s, 6H), 1.77 (s, 18H), 1.97 (d, ³*J* = 14.7 Hz, 2H), 3.13 (s, 2H), 3.21 (d, ³*J* = 13.6 Hz, 2H), 6.97 (d, ³*J* = 8.4 Hz, 2H), 7.65 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.6 Hz, 2H), 8.76 (d, ³*J* = 4.3 Hz, 2H), 9.03 (d, ³*J* = 4.5 Hz, 2H), 9.12 (d, ⁴*J* = 2.6 Hz, 2H), 9.22 (d, ³*J* = 4.4 Hz, 2H), 9.27 (d, ³*J* = 4.3 Hz, 2H), 9.99 (s, 2H). ¹³C NMR data are not available due to decomposition during the measurement. MS (MALDI): *m/z* 758.207 [M]^{•+}. HRMS (ESI, positive) calcd for C₅₀H₄₆MgN₄O₂ [M]⁺ 758.34712, found 758.34657.

BHP Cu-10d (strap: 2Me, *t*-Bu-aryl; *meso*: $2 \times H$) prepared from **10d** (73 mg). 76 mg, 98% yield. mp >300 °C. MS (MALDI): *m/z* 797.254 [M] ^{•+}. HRMS (ESI): calcd for $C_{50}H_{46}CuN_4O_2$ [M + H]⁺ 797.29168, found 797.29113.

BHP In-10d (strap: 2Me, *t*-Bu-aryl; *meso*: 2 × H) prepared from **10d** (73 mg). 88 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.83 (s, 6H), 1.81 (s, 18H), 1.98 (d, ³*J* = 13.0 Hz, 2H), 3.10 (s, 2H), 3.23 (d, ³*J* = 13.0 Hz, 2H), 7.01 (d, ³*J* = 8.3 Hz, 2H), 7.74 (dd, ³*J* = 8.3 Hz, ⁴*J* = 2.5 Hz, 2H), 8.95 (d, ³*J* = 4.6 Hz, 2H), 9.18 (d, ³*J* = 4.6 Hz, 2H), 9.22 (d, ⁴*J* = 2.5 Hz, 2H), 9.45 (d, ³*J* = 4.6 Hz, 2H), 9.51 (d, ³*J* = 4.6 Hz, 2H), 10.31 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.1, 32.1, 35.0, 71.6, 105.7, 115.8, 120.4, 120.7, 126.1, 127.0, 128.5, 130.1, 132.4, 133.0, 133.2, 133.7, 134.3, 146.1, 148.4, 148.5, 149.9, 151.1, 158.4 ppm. MS (MALDI): *m/z* 884.254 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₆ClInN₄O₂ [M + H]⁺ 884.23427, found 884.23358.

BHP V-10d (strap: 2Me, *t*-Bu-aryl; *meso*: $2 \times H$) prepared from 10d (73 mg). 80 mg, 99% yield. mp >300 °C. MS (MALDI): *m/z* 801.323 [M]^{•+}. HRMS (ESI): calcd for $C_{50}H_{46}N_4O_3V$ [M]⁺ 801.30096, found 801.30041.

BHP Zn-10e (strap: 4Me, *t*-Oct-aryl; *meso*: 2 × H) prepared from **10e** (75 mg). 79 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –0.80 (s, 6H), 1.78 (s, 18H), 1.99 (d, ³*J* = 13.4 Hz, 2H), 3.22 (s, 2H), 3.27 (d, ³*J* = 13.4 Hz, 2H), 7.01 (d, ³*J* = 8.4 Hz, 2H), 7.68 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.6 Hz, 2H), 8.91 (d, ³*J* = 4.5 Hz, 2H), 9.12 (d, ⁴*J* = 2.6 Hz, 2H), 9.19 (d, ³*J* = 4.5 Hz, 2H), 9.29 (d, ³*J* = 4.5 Hz, 2H), 9.33 (d, ³*J* = 4.5 Hz, 2H), 10.06 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 1.2, 14.3, 29.9, 32.2, 35.0, 72.0, 105.7, 113.6, 120.5, 120.9, 126.2, 126.4, 127.9, 130.4, 131.7, 132.3, 132.9, 132.9, 135.1, 146.1, 148.9, 149.0, 149.5, 151.1, 159.0 ppm. MS (MALDI): *m/z* 798.315 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₆N₄O₂Zn [M]⁺ 798.29122, found 798.29111.

BHP In-10f (strap: 2Me, *t*-Oct-aryl; *meso*: $2 \times Br$) prepared from 10f (89 mg). 79 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.80 (s, 6H), 1.22 (s, 18H), 1.88 (d, ³*J* = 11.2 Hz, 12H), 2.02 (d, ³*J* = 13.4 Hz, 2H), 2.19 (d, ³*J* = 14.5 Hz, 2H), 2.28 (d, ³*J* = 14.5 Hz, 2H), 3.12 (s, 2H), 3.26 (d, ³*J* = 13.4 Hz, 2H), 7.03 (d, ³*J* = 8.3 Hz, 2H), 7.75 (dd, ³*J* = 8.3 Hz, ⁴*J* = 2.5 Hz, 2H), 8.94 (d, ³*J* = 4.5 Hz, 2H), 9.20 (d, ³*J* = 4.5 Hz, 2H), 9.25 (d, ⁴*J* = 2.5 Hz, 2H), 9.47 (d, ³*J* = 4.6 Hz, 2H), 9.52 (d, ³*J* = 4.5 Hz, 2H), 10.33 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.4, 31.9, 32.5, 33.0, 38.9, 57.5, 71.6, 105.7, 115.8, 120.3, 120.7, 126.1, 127.9, 129.3, 130.1, 132.3, 132.9, 133.1, 133.7, 134.1, 144.9, 148.3, 148.4, 149.9, 151.1, 158.3 ppm. MS (MALDI): *m/z* 996.339 [M]^{•+}. HRMS (ESI): calcd for C₅₈H₆₂InN₄O₂ [M - Cl]⁺ 961.39061, found 961.39169.

BHP In-10h (strap: 2Br, *t*-Oct-aryl; *meso*: 2 × H) prepared from **10h** (98 mg). 112 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 1.19 (s, 18H), 1.85 (d, ³J = 11.3 Hz, 12H), 2.12 (s, 6H), 2.16 (d, ³J = 14.6 Hz, 2H), 2.26 (d, ³J = 14.6 Hz, 2H), 3.18 (d, ³J = 14.4 Hz, 2H), 3.50 (s, 2H), 7.02 (d, ³J = 8.4 Hz, 2H), 7.74 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 8.87 (d, ³J = 4.5 Hz, 2H), 9.17 (d, ³J = 4.5 Hz, 2H), 9.24 (d, ⁴J = 2.5 Hz, 2H), 9.49 (d, ³J = 4.5 Hz, 2H), 9.54 (d, ³J = 4.5 Hz, 2H), 10.40 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 20.9, 31.8, 32.5, 33.0, 33.0, 38.9, 57.5, 72.6, 106.8, 114.2, 121.0, 124.7, 128.0, 129.7, 132.0, 132.7, 133.0, 133.4, 134.1, 134.8, 145.7, 148.7, 149.0, 149.7, 150.5, 157.8, 177.3 ppm. MS (MALDI): *m/z* 1124.111 [M]^{•+}. HRMS (ESI): calcd for C₅₆H₅₆Br₂ClInN₄NaO₂ [M + Na]⁺ 1147.13895, found 1147.13764.

BHP Ni-10i (strap: 4H, *t*-Oct-aryl; *meso*: 2 × H) prepared from **10i** (83 mg). 87 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 1.10 (s, 18H), 1.76 (s, 12H), 2.11 (s, 4H), 3.75 (s, 4H), 4.11 (s, 4H), 6.76 (d, ³*J* = 8.4 Hz, 2H), 7.60 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 8.83 (d, ³*J* = 4.7 Hz, 4H), 8.91 (d, ⁴*J* = 2.5 Hz, 2H), 9.02 (d, ³*J* = 4.7 Hz, 4H), 9.60 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.8, 142.6, 142.3, 141.0, 134.8, 132.9, 132.0, 130.0, 129.0, 127.5, 125.2, 114.4, 112.2, 104.2, 71.2, 57.5, 38.7, 32.9, 32.4 ppm. MS (MALDI): *m*/*z* 876.402 [M]^{•+}. HRMS (ESI): calcd for C₅₆H₅₈N₄NiO₂ [M]⁺ 876.39078, found 876.39185.

BHP Zn-11a (strap: 4Me, H-aryl; *meso*: H/(4-*t*-Bu)Ph) prepared from 11a (80 mg). 84 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –0.15 (s, 6H), –0.02 (s, 6H), 1.62 (s, 9H), 3.52 (d, ³J = 1.2 Hz, 4H), 6.59–6.67 (m, 2H), 7.54–7.62 (m, 4H), 7.74 (d, ³J = 8.0, 2H), 7.99–8.18 (m, 2H), 8.82–8.87 (m, 4H), 8.93–8.98 (m, 4H), 9.19 (d, ³J = 4.4, 2H), 9.92 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 158.8, 151.4, 151.3, 150.3, 149.5, 148.9, 140.2, 134.4, 132.7, 132.5, 132.2, 132.2, 131.4, 131.3, 131.1, 130.7, 130.0, 127.6, 123.8, 121.4, 119.8, 115.2, 111.1, 104.6, 63.8, 35.2, 32.1, 13.5 ppm. MS (MALDI): m/z 846.343 [M]^{•+}. HRMS (ESI): calcd for C₅₄H₄₆N₄O₂Zn [M]⁺ 846.29067, found 846.29060.

BHP Cu-11a (strap: 4Me, H-aryl; meso: H/(4-t-Bu)Ph) prepared from 11a (80 mg). 84 mg, 97% yield. mp >300 °C. MS (MALDI): m/

z 845.315 $[M]^{\bullet+}$. HRMS (ESI): calcd for $C_{54}H_{46}N_4O_2Cu$ $[M]^+$ 845.29168, found 845.29225.

BHP Ni-11a (strap: 4Me, H-aryl; *meso*: H/(4+*t*-Bu)Ph) prepared from 11a (80 mg). 83 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.41 (s, 6H), 0.69 (s, 6H), 1.52 (s, 9H), 4.11 (d, ³*J* = 9.8 Hz, 2H), 4.21 (d, ³*J* = 9.8 Hz, 2H), 6.81 (dd, ³*J* = 8.1 Hz, ⁴*J* = 1.1 Hz, 2H), 7.44–7.50 (m, 2H), 7.53–7.60 (m, 2H), 7.63–7.67 (m, 2H), 7.87 (d, ³*J* = 7.8 Hz, 2H), 8.66 (d, ³*J* = 4.9 Hz, 2H), 8.68 (d, ³*J* = 4.7 Hz, 2H), 8.78 (dd, ³*J* = 4.7 Hz, ⁴*J* = 1.0 Hz, 2H), 8.88 (dd, ³*J* = 4.7 Hz, ⁴*J* = 1.0 Hz, 2H), 9.40 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.3, 14.9, 29.9, 31.8, 35.0, 63.9, 77.4, 103.0, 110.7, 119.1, 119.4, 124.0, 129.0, 129.9, 130.2, 131.5, 131.8, 131.8, 132.4, 132.6, 133.5, 140.4, 141.1, 141.6, 142.0, 158.8 ppm. MS (MALDI): *m*/*z* 840.284 [M]^{•+}. HRMS (ESI): calcd for C₅₄H₄₆N₄NiO₂ [M + H]⁺ 840.29742, found 840.29714.

BHP Pd-11b (strap: 4Me, *t*-Bu-aryl; *meso*: H/(4-*t*-Bu)Ph) prepared from **11b** (90 mg). 100 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.07 (s, 6H), 0.08 (s, 6H), 1.59 (s, 9H), 1.74 (s, 18H), 3.68 (d, ³*J* = 10.2 Hz, 2H), 3.71 (d, ³*J* = 10.2 Hz, 2H), 6.63 (d, ³*J* = 8.4 Hz, 2H), 7.60 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 7.2 (s, 2H), 8.71 (d, ³*J* = 4.8 Hz, 2H), 8.77 (d, ³*J* = 4.8 Hz, 2H), 8.90 (d, ³*J* = 4.8 Hz, 2H), 8.96 (d, ⁴*J* = 2.5 Hz, 2H), 9.04 (d, ³*J* = 4.8 Hz, 2H), 9.84 (s, 1H) ppm. ¹³C NMR (150 MHz, CDCl3): δ 13.5, 13.6, 31.8, 32.2, 34.8, 64.3, 105.2, 110.8, 115.9, 121.5, 123.7, 125.4, 126.2, 130.6, 130.8, 131.0, 131.0, 131.1, 131.3, 133.9, 138.9, 139.9, 140.4, 141.9, 142.3, 142.4, 150.4, 156.6 ppm. MS (MALDI): *m/z* 1000.310 [M]^{*+}. HRMS (ESI): calcd for C₆₂H₆₂N₄NaO₂Pd [M + Na]⁺ 1023.37998, found 1023.38267.

BHP Ni-11b (strap: 4Me, *t*-Bu-aryl; *meso*: H/(4-*t*-Bu)Ph) prepared from **11b** (90 mg). 96 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.40 (s, 6H), 0.70 (s, 6H), 1.54 (s, 9H), 1.70 (s, 18H), 4.09 (d, ³*J* = 9.8 Hz, 2H), 4.20 (d, ³*J* = 9.8 Hz, 2H), 6.75 (d, ³*J* = 8.5 Hz, 2H), 7.60 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 7.63–7.68 (m, 2H), 7.88 (d, ³*J* = 7.7 Hz, 2H), 8.66 (d, ³*J* = 4.9 Hz, 2H), 8.68 (d, ³*J* = 4.9 Hz, 2H), 8.79 (d, ³*J* = 4.7 Hz, 2H), 8.86 (d, ⁴*J* = 2.5 Hz, 2H), 8.88 (d, ³*J* = 4.7 Hz, 2H), 9.38 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.7, 150.6, 142.2, 142.2, 141.8, 141.3, 140.6, 137.9, 133.7, 132.7, 132.7, 132.5, 132.5, 132.1, 132.0, 131.8, 128.6, 127.9, 126.3, 124.2, 119.2, 112.7, 110.2, 103.2, 64.2, 35.1, 34.9, 32.4, 32.1, 32.0, 15.0, 14.5 ppm. MS (MALDI): *m*/*z* 952.392 [M]^{•+}. HRMS (ESI): calcd for C₆₂H₆₂N₄NaNiO₂ [M + Na]⁺ 975.41185, found 975.41241.

BHP Cu-11b (strap: 4Me, *t*-Bu-aryl; *meso*: H/(4-*t*-Bu)Ph) prepared from **11b** (90 mg). 95 mg, 98% yield. mp >300 °C. MS (MALDI): m/z 957.399 [M]^{•+}. HRMS (ESI): calcd for C₆₂H₆₂CuN₄O₂ [M + H]⁺ 957.41633, found 957.41400.

BHP Zn-11b (strap: 4Me, *t*-Bu-aryl; *meso*: H/(4-*t*-Bu)Ph) prepared from **11b** (90 mg). 95 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.15 (s, 6H), -0.04 (s, 6H), 1.61 (s, 9H), 1.76 (s, 18H), 3.51 (s, 4H), 6.56 (d, ³J = 8.4 Hz, 2H), 7.59 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 8.08 (s, 2H), 8.84 (d, ³J = 4.6 Hz, 2H), 8.86 (d, ³J = 4.6 Hz, 2H), 8.86 (d, ³J = 4.5 Hz, 2H), 9.92 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 1.2, 13.2, 13.3, 31.9, 32.3, 34.8, 35.0, 64.1, 104.5, 110.7, 115.7, 123.6, 125.0, 130.6, 130.7, 130.8, 131.3, 131.9, 132.1, 132.2, 132.3, 134.3, 140.0, 142.4, 148.8, 149.2, 150.2, 151.3, 151.3, 156.6 ppm. MS (MALDI): m/z 958.419 [M]^{•+}. HRMS (ESI): calcd for C₆₂H₆₃N₄O₂Zn [M + H]⁺ 959.42370, found 959.42494.

BHP Zn-11e (strap: 4Me, H-aryl; *meso*: H/*n*-Bu) prepared from **11e** (72 mg). 76 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –0.14 (s, 6H), –0.04 (s, 6H), 1.15 (t, ³*J* = 7.4 Hz, 3H), 1.83 (q, ³*J* = 7.4 Hz, 2H), 2.55 (p, ³*J* = 7.9 Hz, 2H), 3.53 (d, ³*J* = 10.0 Hz, 2H), 3.58 (d, ³*J* = 10.0 Hz, 2H), 4.89 (t, ³*J* = 7.3 Hz, 2H), 6.60– 6.67 (m, 2H), 7.55–7.62 (m, 4H), 8.90 (dd, ³*J* = 4.5 Hz, ⁴*J* = 2.2 Hz, 4H), 8.93–8.98 (m, 2H), 9.14 (d, ³*J* = 4.5 Hz, 2H), 9.39 (d, ³*J* = 4.6 Hz, 2H), 9.83 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 158.9, 150.9, 150.7, 149.6, 148.5, 132.7, 132.1, 132.1, 131.5, 131.3, 131.1, 130.7, 129.9, 129.3, 127.7, 121.9, 119.8, 114.7, 111.1, 104.1, 77.0, 63.8, 41.8, 35.6, 30.1, 24.1, 14.6, 13.5, 13.5 ppm. MS (MALDI): *m/z* 770.241 [M]^{•+}. HRMS (ESI): calcd for C₄₈H₄₃N₄O₂Zn [M + H]⁺ 771.26720, found 771.26606. **BHP Ni-11e** (strap: 4Me, H-aryl; *meso*: H/*n*-Bu) prepared from **11e** (72 mg). 78 mg, 97% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.49 (s, 6H), 0.52 (s, 6H), 0.89–0.93 (m, 4H), 1.40 (m, 2H), 2.04–2.12 (m, 2H), 4.17 (s, 4H), 4.53 (app t, ^{app}J = 7.9 Hz, 2H), 6.82 (dd, ³J = 0.7 Hz, ⁴J = 8.1 Hz, 2H), 7.48 (app dt, ⁴J = 1.0 Hz, ^{app}J = 7.6 Hz, 2H), 7.58 (app dt, ⁴J = 1.7 Hz, ^{app}J = 7.9 Hz, 2H), 8.73 (d, ³J = 5.0 Hz, 2H), 8.75 (d, ³J = 4.8 Hz, 2H), 8.78 (dd, ³J = 7.3 Hz, ⁴J = 1.8 Hz, 2H), 8.82 (d, ³J = 4.8 Hz, 2H), 9.14 (d, ³J = 5.0 Hz, 2H), 9.32 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.0, 14.3, 14.5, 14.6, 23.2, 29.9, 31.7, 31.8, 32.1, 33.3, 38.9, 64.0, 102.6, 110.8, 111.7, 118.9, 119.4, 129.0, 129.9, 130.1, 130.3, 131.4, 132.0, 132.2, 132.4, 140.1, 141.3, 141.3, 158.8 ppm. MS (MALDI): *m*/*z* 764.142 [M]*+ 1RMS (ESI, positive) calcd for C₄₈H₄₂N₄NaNiO₂ [M + Na]⁺ 787.25535, found 787.25509.

BHP Zn-11i (strap: 4Me, *t*-Bu-aryl; *meso*: H/*n*-Bu) prepared from 11i (82 mg). 88 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ –0.19 (s, 6H), –0.10 (s, 6H), 1.14 (t, ³*J* = 7.4 Hz, 3H), 1.79 (s, 18H), 1.83 (m, 2H), 2.51 (h, ³*J* = 8.0 Hz, 7.3, 2H), 3.47 (d, ³*J* = 10.4 Hz, 2H), 3.51 (d, ³*J* = 10.2 Hz, 2H), 4.82 (t, ³*J* = 8.1 Hz, 2H), 6.55 (d, ³*J* = 8.3 Hz, 2H), 7.58 (dd, ³*J* = 8.3 Hz, ⁴*J* = 2.5 Hz, 2H), 8.84 (d, ³*J* = 4.5 Hz, 2H), 8.86 (d, ³*J* = 4.5 Hz, 2H), 9.01 (d, ⁴*J* = 2.4 Hz, 2H), 9.11 (d, ³*J* = 4.4, 2H), 9.32 (d, ³*J* = 4.6 Hz, 2H), 9.78 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 156.7, 150.8, 150.6, 149.5, 148.4, 142.5, 132.2, 131.9, 131.6, 131.1, 131.0, 130.7, 129.1, 126.0, 125.2, 121.7, 115.1, 110.6, 104.0, 64.1, 41.7, 35.5, 35.0, 32.5, 30.1, 24.0, 14.6, 13.4 ppm. MS (MALDI): *m*/z 882.413 [M]*+. HRMS (ESI): calcd for C₅₆H₅₉N₄O₂Zn [M + H]⁺ 883.39240, found 883.39227.

BHP Zn-11h (strap: 2Me, H-aryl; meso: H/n-Bu) prepared from 11h (70 mg). 749 mg, 97% yield. mp >300 °C. ¹H NMR (400 MHz, $CDCl_3$): $\delta -0.78$ (s, 6H), 1.16 (t, ${}^{3}J = 7.4$ Hz, 3H), 1.84 (d, ${}^{3}J = 7.4$ Hz, 2H), 2.06 (d, ³*J* = 14.1 Hz, 1H), 2.11 (d, ³*J* = 14.1 Hz, 1H), 2.48-2.59 (m, 2H), 3.27–3.34 (m, 4H), 4.64–4.80 (m, 2H), 7.07 (dd, ${}^{3}J$ = 8.0 Hz, ${}^{4}I = 1.0, 1$ H), 7.08 (dd, ${}^{3}I = 8.0$ Hz, ${}^{4}I = 1.0$ Hz, 1H), 7.66 (app dt, ${}^{4}J = 1.8$ Hz, ${}^{app}J = 7.8$ Hz, 2H), 7.79 (app tt, ${}^{3}J = 1.3$ Hz, ${}^{app}J = 7.8$ Hz, 2H), 8.72 (d, ${}^{3}J$ = 4.6 Hz, 1H), 8.74 (d, ${}^{3}J$ = 4.6 Hz, 1H), 8.98 (app dt, ${}^{4}J$ = 1.8 Hz, ${}^{app}J$ = 7.6 Hz, 2H), 9.02 (d, ${}^{3}J$ = 4.6 Hz, 1H), 9.04 $(d, {}^{3}J = 4.6 \text{ Hz}, 1\text{H}), 9.13 (d, {}^{3}J = 3.2 \text{ Hz}, 1\text{H}), 9.14 (d, {}^{3}J = 3.2 \text{ Hz},$ 1H), 9.31 (d, ${}^{3}J$ = 4.6 Hz, 1H), 9.34 (d, ${}^{3}J$ = 4.6 Hz, 1H), 9.76 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.3, 14.7, 24.0, 35.3, 41.7, 53.6, 71.9, 72.1, 104.5, 112.8, 120.6, 120.7, 121.6, 121.7, 122.0, 123.2, 123.2, 126.3, 126.3, 129.6, 129.8, 129.8, 129.8, 130.2, 130.4, 130.6, 130.8, 130.8, 131.5, 132.0, 132.1, 132.6, 132.6, 135.9, 136.0, 148.3, 148.4, 148.6, 148.9, 149.4, 149.4, 149.9, 150.4, 161.1, 161.3 ppm. MS (MALDI, positiv): m/z 742.253 [M]^{•+}. HRMS (ESI): calcd for C₄₆H₃₈N₄O₂Zn [M]⁺ 742.22807, found 742.22823.

BHP Zn-11g (strap: 4Me, H-aryl; *meso*: H/*n*-Hex) prepared from 11g (74 mg). 79 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.04 (s, 6H), -0.00 (s, 6H), 0.95 (t, J = 7.3 Hz, 3H), 1.41 (m, 2H), 1.48–1.58 (m, 2H), 1.78–1.88 (m, 2H), 2.52–2.62 (m, 2H), 3.60 (dd, J = 10.1 Hz, 4H), 4.85–4.92 (m, 2H), 6.62–6.69 (m, 2H), 7.54–7.63 (m, 4H), 8.89 (d, J = 4.5 Hz, 4H), 8.93–8.97 (m, 2H), 9.14 (d, J = 4.5 Hz, 2H), 9.39 (d, J = 4.6 Hz, 2H), 9.82 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 13.2, 13.3, 14.4, 22.9, 29.9, 30.5, 32.1, 35.8, 39.6, 63.7, 111.0, 114.7, 119.6, 121.8, 127.5, 129.1, 129.7, 130.5, 130.9, 131.0, 131.4, 131.8, 132.0, 132.6, 148.5, 149.5, 150.6, 150.8, 158.7 ppm. MS (MALDI): *m*/*z* 798.215 [M]^{•+}. HRMS (ESI, positive) calcd for C₅₀H₄₇N₄O₂Zn [M + H]⁺ 799.29850, found 799.29746.

BHP Ni-11g (strap: 4Me, H-aryl; *meso*: H/*n*-Hex) prepared from **11g** (74 mg). 79 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.50 (s, 6H), 0.53 (s, 6H), 1.26 (m, 6H), 1.43 (m, 2H), 2.05–2.12 (m, 2H), 4.16 (s, 4H), 4.52 (app t, ^{app}J = 8.0 Hz, 2H), 6.82 (dd, ³J = 8.1 Hz, ⁴J = 0.8 Hz, 2H), 7.49 (app dt, ⁴J = 1.1 Hz, ^{app}J = 7.6 Hz, 2H), 7.59 (app dt, ⁴J = 1.8 Hz, ^{app}J = 7.9 Hz, 2H), 8.74 (dd, ³J = 4.9 Hz, ⁴J = 0.8 Hz, 4H), 8.77 (dd, ³J = 7.3 Hz, ⁴J = 1.8 Hz, 2H), 8.86 (d, ³J = 4.8 Hz, 2H), 9.13 (d, ³J = 4.9 Hz, 2H), 9.35 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.6, 14.6, 29.9, 33.7, 36.9, 64.0, 102.6, 11.8, 111.7, 119.0, 119.3, 129.0, 129.9, 130.1, 130.3, 131.4, 132.0, 132.2, 132.4, 140.1, 141.3, 141.3, 158.8 ppm. MS (MALDI): *m/z* 792.223 [M]^{•+}. HRMS (ESI): calcd for C₅₀H₄₆N₄NiNaO₂ [M + Na]⁺ 815.28665, found 815.28512.

BHP Zn-12a (strap: 4Me, H-aryl; *meso*: $2 \times (4-t$ -Bu)Ph) prepared from **12a** (92 mg). 97 mg, 99% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.02 (s, 12H), 1.61 (s, 18H), 3.60 (s, 4H), 6.61–6.68 (m, 2H), 7.53–7.63 (m, 4H), 7.74 (d, ³J = 7.9 Hz, 4H), 8.08 (br. s, 4H), 8.82 (d, ³J = 4.6 Hz, 4H), 8.85 (d, ³J = 4.6 Hz, 4H), 8.89–8.96 (m, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 159.0, 151.3, 150.4, 149.6, 140.2, 134.4, 133.1, 132.5, 131.4, 131.2, 130.8, 129.9, 127.5, 123.8, 119.9, 111.4, 77.7, 77.6, 77.4, 77.1, 77.0, 64.1, 35.2, 32.1, 30.1, 13.5, -2.8, -2.9 ppm. MS (MALDI): *m/z* 978.355 [M]^{•+}. HRMS (ESI): calcd for C₆₄H₅₉N₄O₂Zn [M + H]⁺ 979.39240, found 979.39141.

BHP Ni-12a (strap: 4Me, H-aryl; *meso*: 2 × (4-*t*-Bu)Ph) prepared from **12a** (90 mg). 97 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.66 (s, 12H), 1.54 (s, 18H), 4.22 (s, 4H), 6.69–6.77 (m, 2H), 7.35–7.41 (m, 4H), 7.66 (d, ³*J* = 8.6 Hz, 4H), 7.87 (d, ³*J* = 7.7 Hz, 4H), 8.65 (d, ³*J* = 4.9 Hz, 4H), 8.69 (d, ³*J* = 4.9 Hz, 4H), 8.72–8.75 (m, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.7, 31.8, 35.0, 64.0, 110.7, 119.3, 124.1, 128.9, 129.9, 130.3, 131.6, 131.9, 132.2, 132.6, 133.4, 137.6, 140.8, 141.5, 150.4, 158.7 ppm. MS (MALDI): *m*/*z* 972.399 [M]^{•+}. HRMS (ESI): calcd for C₆₄H₅₉N₄NiO₂ [M + H]⁺ 973.39860, found 973.39745.

BHP In-13h (strap: 2Br, *t*-Oct-aryl; *meso*: 2 × Br) prepared from **13h** (113 mg). 126 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 1.15 (s, 18H), 1.81 (d, ³J = 7.3 Hz, 12H), 2.05 (d, ³J = 14.5 Hz, 2H), 2.12 (d, ³J = 14.7 Hz, 2H), 2.21 (d, ³J = 14.7 Hz, 2H), 3.31 (d, ³J = 14.5 Hz, 2H), 3.77 (s, 2H), 7.04 (d, ³J = 8.4 Hz, 2H), 7.73 (dd, ³J = 8.4 Hz, ⁴J = 2.5 Hz, 2H), 8.71 (d, ³J = 4.7 Hz, 2H), 9.03 (d, ³J = 4.8 Hz, 2H), 9.07 (d, ⁴J = 2.5 Hz, 2H), 9.73 (d, ³J = 4.8 Hz, 2H), 9.07 (d, ⁴J = 2.5 Hz, 2H), 9.73 (d, ³J = 4.8 Hz, 2H), 9.07 (d, ⁴J = 2.5 Hz, 2H), 9.73 (d, ³J = 4.8 Hz, 2H), 9.81 (d, ³J = 4.8 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 31.8, 32.5, 33.0, 33.0, 38.9, 57.4, 72.9, 105.8, 114.6, 117.0, 121.2, 125.1, 128.3, 129.6, 133.1, 133.4, 133.5, 133.9, 135.0, 136.4, 145.8, 149.4, 149.6, 150.0, 150.8, 157.7 ppm. MS (MALDI): *m*/z 1282.052 [M]*+ 1RMS (ESI): calcd for C₅₆H₅₄Br₄ClInN₄NaO₂ [M + Na]+ 1302.95998, found 1302.96124.

BHP In-13f (strap: 2Me, *t*-Oct-aryl; *meso*: 2 × Br) prepared from **13f** (101 mg). 114 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ -0.58 (s, 6H), 1.16 (s, 18H), 1.82 (d, ³*J* = 7.1 Hz, 12H), 2.11–2.23 (m, 6H), 3.28–3.43 (m, 4H), 7.02 (d, ³*J* = 8.4 Hz, 2H), 7.73 (dd, ³*J* = 8.4 Hz, ⁴*J* = 2.5 Hz, 2H), 8.76 (d, ³*J* = 4.8 Hz, 2H), 9.05 (d, ³*J* = 4.8 Hz, 2H), 9.07 (d, ⁴*J* = 2.5 Hz, 2H), 9.72 (d, ³*J* = 4.8 Hz, 2H), 9.07 (d, ⁴*J* = 2.5 Hz, 2H), 9.72 (d, ³*J* = 4.8 Hz, 2H), 9.07 (d, ⁴*J* = 2.5 Hz, 2H), 9.72 (d, ³*J* = 4.8 Hz, 2H), 9.07 (d, ⁴*J* = 2.5 Hz, 2H), 9.72 (d, ³*J* = 4.8 Hz, 2H), 9.79 (d, ³*J* = 4.8 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.5, 31.9, 32.5, 33.0, 33.0, 38.9, 57.5, 71.8, 104.6, 118.1, 120.4, 121.1, 126.6, 128.3, 129.2, 130.5, 133.4, 133.5, 134.1, 134.6, 135.3, 145.0, 148.8, 149.1, 150.2, 151.3, 158.2 ppm. MS (MALDI): *m/z* 1152.152 [M]^{•+}. HRMS (ESI): calcd for C₅₈H₆₀Br₂ClInN₄NaO₂ [M + Na]⁺ 1175.17025, found 1175.16983.

BHP Ni-13c (strap: 4Me, *t*-Bu-aryl; *meso*: $2 \times Br$) prepared from **13c** (93 mg). 97 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.60 (s, 12H), 1.52(s, 2H), 1.69 (s, 18H), 4.18 (s, 4H), 6.76 (d, ³J = 8.5 Hz, 2H), 7.61 (dd, ³J = 8.5 Hz, ⁴J = 2.5 Hz, 2H), 8.64 (d, ³J = 4.9 Hz, 4H), 8.77 (d, ⁴J = 2.5 Hz, 2H), 9.17 (d, ³J = 4.9 Hz, 4H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.3, 14.8, 31.8, 32.2, 34.7, 64.1, 77.4, 110.2, 114.5, 126.6, 127.5, 127.8, 131.6, 132.1, 133.5, 133.6, 140.9, 142.2, 142.3, 156.4 ppm. MS (MALDI,): *m/z* 975.931 [M]^{•+}. HRMS (ESI): calcd for C₅₂H₄₈Br₂N₄NaNiO₂ [M + Na]⁺ 999.13897, found 999.14153.

BHP Ni-14d (strap: 4Me, *t*-Bu-aryl; *meso*: H/Br) prepared from 14d (85 mg). 89 mg, 98% yield. mp >300 °C. ¹H NMR (400 MHz, CDCl₃): δ 0.33 (s, 6H), 0.70 (s, 6H), 1.72 (s, 18H), 4.09 (d, ³*J* = 9.8 Hz, 2H), 4.19 (d, ³*J* = 9.8 Hz, 2H), 6.75 (d, ³*J* = 8.5 Hz, 2H), 7.62 (dd, ³*J* = 8.5 Hz, ⁴*J* = 2.5 Hz, 2H), 8.71 (d, ³*J* = 4.7 Hz, 2H), 8.74 (d, ³*J* = 4.9 Hz, 2H), 8.81 (d, ³*J* = 4.7 Hz, 2H), 8.83 (d, ⁴*J* = 2.5 Hz, 2H), 9.27 (d, ³*J* = 4.9 Hz, 2H), 9.34 (s, 1H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ 14.4, 14.8, 32.2, 34.7, 64.0, 77.4, 101.1, 103.9, 110.1, 113.2, 126.4, 127.6, 128.0, 131.5, 131.9, 132.2, 132.6, 132.9, 133.1, 133.3, 140.2, 141.4, 142.1, 142.2, 142.2, 156.5 ppm. MS (MALDI): *m/z* 898.172 [M]^{•+}. HRMS (ESI): calcd for C₅₂H₄₉BrN₄NaNiO₂ [M + Na]⁺ 921.22846, found 921.22673.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b02259.

Crystallographic data for In-10d (CIF)

Crystallographic data for V-10d (CIF)

¹H and ¹³C NMR, NOESY, and COSY for 10f,h, chromatographic details for the enantiomeric resolution of 10d,f,h, 11h, 13b,d,f, 14a, Zn-10b, Zn-10d, Ni-10d, Pd-10d, Cu-10d, Pd-10d, Mg-10d, V-10d, In-10d, and In-13f, UV/vis spectra, additional information on the crystallographic measurements, and Cartesian coordinates and energies of the computationally investigated compounds (PDF)

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notes

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REFERENCES

(1) Handbook of Porphyrin Science; Kadish, K. M., Smith, K. M., Guilard, R., Eds; World Scientific: Singapore, 2010; Vols. 1-20.

(2) Dolmans, D. E. J. G. J.; Fukumura, D.; Jain, R. K. Nat. Rev. Cancer 2003, 3, 380.

(3) *The Porphyrins*; Dolphin, D., Ed., Academic Press: New York, 1979; Vol. 6 (Biochemistry, Part A).

(4) Senge, M. O.; Fazekas, M.; Notaras, E. G. A.; Blau, W. J.; Zawadzka, M.; Locos, O. B.; Ni Mhuircheartaigh, E. M. Adv. Mater. (Weinheim, Ger.) 2007, 19, 2737.

(5) Senge, M. O. Chem. Commun. 2011, 47, 1943.

(6) Ryan, A.; Gehrold, A.; Perusitti, R.; Pintea, M.; Fazekas, M.; Locos, O. B.; Blaikie, F.; Senge, M. O. *Eur. J. Org. Chem.* 2011, 2011, 5817.

(7) Aratani, N.; Kim, D.; Osuka, A. Acc. Chem. Res. 2009, 42, 1922.

(8) Wytko, J. A.; Graf, E.; Weiss, J. J. Org. Chem. 1992, 57, 1015.

(9) Baldwin, J. E.; Klose, T.; Peters, M. J. Chem. Soc., Chem. Commun. 1976, 881.

(10) Osuka, A.; Kobayashi, F.; Maruyama, K. Bull. Chem. Soc. Jpn. 1991, 64, 1213.

(11) Wang, Q. M.; Bruce, D. W. Tetrahedron Lett. 1996, 37, 7641.

(12) Gehrold, A. C.; Bruhn, T.; Schneider, H.; Radius, U.; Bringmann, G. Org. Lett. 2015, 17, 210.

(13) Koepf, M.; Conradt, J.; Szmytkowski, J.; Wytko, J. A.; Allouche, L.; Kalt, H.; Balaban, T. S.; Weiss, J. *Inorg. Chem.* **2011**, *50*, 6073.

(14) Takeuchi, M.; Fujikoshi, C.; Kubo, Y.; Kaneko, K.; Shinkai, S. Angew. Chem., Int. Ed. 2006, 45, 5494.

(15) Zhou, Z.; Shen, M.; Cao, C.; Liu, Q.; Yan, Z. Chem. - Eur. J. 2012, 18, 7675.

(16) Zhou, Z.; Cao, C.; Liu, Q.; Jiang, R. Org. Lett. 2010, 12, 1780.
(17) Fazio, M. A.; Durandin, A.; Tkachenko, N. V.; Niemi, M.;

Lemmetyinen, H.; Schuster, D. I. Chem. - Eur. J. 2009, 15, 7698. (18) Collman, J. P.; Decreau, R. A.; Costanzo, S. Org. Lett. 2004, 6,

(19) Somman, J. 1., Decread, R. H., Sostanzo, S. Org, Edit. 2004, 6, 1033.
(19) Shao, X.-B.; Jiang, X.-K.; Wang, X.-Z.; Li, Z.-T.; Zhu, S.-Z.

(19) Shao, A.-D.; Jiang, A.-K.; Wang, A.-Z.; Li, Z.-T.; Zhu, S.-Z. Tetrahedron 2003, 59, 4881.

(20) Note that instead of the formally correct expression "p-xylylene", the commonly found term "p-xylene" is used throughout the paper.

- (21) Phan, D. H. T.; Kim, B.; Dong, V. M. J. Am. Chem. Soc. 2009, 131, 15608.
- (22) Duff, J. C. J. Chem. Soc. 1941, 547.
- (23) Balaban, T. S.; Goddard, R.; Linke-Schaetzel, M.; Lehn, J.-M. J. Am. Chem. Soc. **2003**, 125, 4233.
- (24) Hata, H.; Shinokubo, H.; Osuka, A. J. Am. Chem. Soc. 2005, 127, 8264.
- (25) Brückner, C.; Posakony, J. J.; Johnson, C. K.; Boyle, R. W.; James, B. R.; Dolphin, D. J. Porphyrins Phthalocyanines **1998**, 02, 455.
- (26) Lindsey, J. S. Porphyrin Handbook; Academic Press: New York, 2000; Vol. 1.
- (27) Senge, M. O. Acc. Chem. Res. 2005, 38, 733.
- (28) Kato, A.; Hartnell, R. D.; Yamashita, M.; Miyasaka, H.; Sugiura, K.-i.; Arnold, D. P. J. Porphyrins Phthalocyanines 2004, 08, 1222.
- (29) Sahoo, A. K.; Nakamura, Y.; Aratani, N.; Kim, K. S.; Noh, S. B.; Shinokubo, H.; Kim, D.; Osuka, A. Org. Lett. **2006**, *8*, 4141.
- (30) Hata, H.; Yamaguchi, S.; Mori, G.; Nakazono, S.; Katoh, T.; Takatsu, K.; Hiroto, S.; Shinokubo, H.; Osuka, A. *Chem. - Asian J.* **2007**, *2*, 849.
- (31) Gotz, D. C. G.; Bruhn, T.; Senge, M. O.; Bringmann, G. J. Org. Chem. 2009, 74, 8005.
- (32) Bringmann, G.; Götz, D. C. G.; Gulder, T. A. M.; Gehrke, T. H.; Bruhn, T.; Kupfer, T.; Radacki, K.; Braunschweig, H.; Heckmann, A.; Lambert, C. J. Am. Chem. Soc. **2008**, 130, 17812.
- (33) Götz, D. C. G.; Gehrold, A. C.; Dorazio, S. J.; Daddario, P.; Samankumara, L.; Bringmann, G.; Brückner, C.; Bruhn, T. *Eur. J. Org. Chem.* **2015**, 2015, 3913.
- (34) Blusch, L. K.; Hemberger, Y.; Pröpper, K.; Dittrich, B.; Witterauf, F.; John, M.; Bringmann, G.; Brückner, C.; Meyer, F. *Chem.* - *Eur. J.* **2013**, *19*, 5868.
- (35) Brückner, C.; Götz, D. C. G.; Fox, S. P.; Ryppa, C.; McCarthy, J. R.; Bruhn, T.; Akhigbe, J.; Banerjee, S.; Daddario, P.; Daniell, H. W.; Zeller, M.; Boyle, R. W.; Bringmann, G. J. Am. Chem. Soc. **2011**, 133, 8740.
- (36) Bruhn, T.; Witterauf, F.; Götz, D. C. G.; Grimmer, C. T.; Würtemberger, M.; Radius, U.; Bringmann, G. *Chem. Eur. J.* **2014**, *20*, 3998.
- (37) Bringmann, G.; Götz, D. C. G.; Gulder, T. A. M.; Gehrke, T. H.; Bruhn, T.; Kupfer, T.; Radacki, K.; Braunschweig, H.; Heckmann, A.; Lambert, C. J. Am. Chem. Soc. **2008**, 130, 17812.
- (38) Bruhn, T.; Schaumlöffel, A.; Hemberger, Y.; Bringmann, G. *Chirality* **2013**, *25*, 243.
- (39) Dreuw, A.; Head-Gordon, M. Chem. Rev. 2005, 105, 4009.
- (40) Le Bahers, T.; Adamo, C.; Ciofini, I. J. Chem. Theory Comput. 2011, 7, 2498.
- (41) Lu, T.; Chen, F. J. Comput. Chem. 2012, 33, 580.
- (42) Urbani, M.; Torres, T. Chem. Eur. J. 2014, 20, 16337.
- (43) Crossley, M. J.; Harding, M. M.; Sternhell, S. J. Am. Chem. Soc. 1986, 108, 3608.
- (44) Braun, J.; Koecher, M.; Schlabach, M.; Wehrle, B.; Limbach, H.-H.; Vogel, E. J. Am. Chem. Soc. **1994**, 116, 6593.
- (45) Crossley, M. J.; Field, L. D.; Harding, M. M.; Sternhell, S. J. Am. Chem. Soc. **1987**, 109, 2335.
- (46) Krieger, C.; Dernbach, M.; Voit, G.; Carell, T.; Staab, H. A. Chem. Ber. 1993, 126, 811.
- (47) Neese, F.; Wennmohs, F.; Becker, U.; Bykov, D.; Ganyushin, D.; Hansen, A.; Izsák, R.; Liakos, D. G.; Kollmar, C.; Kossmann, S.; Pantazis, D. A.; Petrenko, T.; Reimann, C.; Riplinger, C.; Roemelt, M.; Sandhöfer, B.; Schapiro, I.; Sivalingam, K.; Wezisla, B. *ORCA, version 3.0.3 ed.*; MPI CEC, Mühlheim a.d.R., Germany, 2014.
- (48) Neese, F. Wiley Interdisciplinary Reviews: Computational Molecular Science 2012, 2, 73.
- (49) Grimme, S.; Ehrlich, S.; Goerigk, L. J. Comput. Chem. 2011, 32, 1456.
- (50) Grimme, S.; Antony, J.; Ehrlich, S.; Krieg, H. J. Chem. Phys. 2010, 132, 154104.
- (51) Izsak, R.; Neese, F. J. Chem. Phys. 2011, 135, 144105.
- (52) Becke, A. D. J. Chem. Phys. 1993, 98, 1372.

- (53) Lee, C.; Yang, W.; Parr, R. G. Phys. Rev. B: Condens. Matter Mater. Phys. **1988**, 37, 785.
- (54) Zhao, Y.; Truhlar, D. G. J. Phys. Chem. A 2005, 109, 5656.
- (55) Tarnopolsky, A.; Karton, A.; Sertchook, R.; Vuzman, D.; Martin, J. M. L. J. Phys. Chem. A **2008**, 112, 3.
- (56) Grimme, S. J. Comput. Chem. 2006, 27, 1787.
- (57) Weigend, F.; Ahlrichs, R. Phys. Chem. Chem. Phys. 2005, 7, 3297.
- (58) Riplinger, C.; Neese, F. J. Chem. Phys. 2013, 138, 034106.
- (59) Riplinger, C.; Sandhoefer, B.; Hansen, A.; Neese, F. J. Chem. Phys. 2013, 139, 134101.
- (60) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Hevd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Keith, T.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, O.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian09, revision D.01 ed.; Gaussian, Inc.: Wallingford, CT, 2013.
- (61) Hattig, C.; Weigend, F. J. Chem. Phys. 2000, 113, 5154.
- (62) Bruhn, T.; Schaumlöffel, A.; Hemberger, Y. SpecDis, version 1.63 ed.; University of Würzburg: Würzburg, Germany, 2015.
- (63) Bringmann, G.; Bruhn, T.; Maksimenka, K.; Hemberger, Y. *Eur. J. Org. Chem.* **2009**, 2009, 2717.
- (64) Guieu, S.; Crane, A. K.; MacLachlan, M. J. Chem. Commun. 2011, 47, 1169.
- (65) Das, S. G.; Doshi, J. M.; Tian, D.; Addo, S. N.; Srinivasan, B.; Hermanson, D. L.; Xing, C. J. Med. Chem. **2009**, *52*, 5937.
- (66) Littler, B. J.; Miller, M. A.; Hung, C.-H.; Wagner, R. W.; O'Shea, D. F.; Boyle, P. D.; Lindsey, J. S. J. Org. Chem. **1999**, 64, 1391.
- (67) Rey, Y. P.; Zimmer, L. E.; Sparr, C.; Tanzer, E.-M.; Schweizer, W. B.; Senn, H. M.; Lakhdar, S.; Gilmour, R. *Eur. J. Org. Chem.* **2014**, 2014, 1202.